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(54) Title: MAMMALIAN TOXICOLOGICAL RESPONSE MARKERS

(57) Abstract: The present invention relates to mammalian nucleic acid and protein molecules comprising a plurality of nucleic acid and protein molecules. The mammalian nucleic acid molecules can be used as hybridizable array elements in a microarray in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action. The protein molecules can be used in a pharmaceutical composition. The present invention also relates to methods for screening compounds and therapeutics for metabolic responses indicative of a toxic compound or molecule.

MAMMALIAN TOXICOLOGICAL RESPONSE MARKERS

This application is filed under the Patent Cooperation Treaty and claims the benefit of U.S. Nonprovisional Application No. 09/443,184, our Docket No. PC-0007 US, filed 19th November, 1999.

TECHNICAL FIELD

The present invention relates to mammalian nucleic acid and protein molecules, and methods for their use in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

BACKGROUND ART

Toxicity testing is a mandatory and time-consuming part of drug development programs in the pharmaceutical industry. A more rapid screen to determine the effects upon metabolism and to detect toxicity of lead drug candidates may be the use of gene expression microarrays. For example, microarrays of various kinds may be produced using full length genes or gene fragments. These arrays can then be used to test samples treated with the drug candidates to elucidate the gene expression pattern associated with drug treatment. This gene pattern can be compared with gene expression patterns associated with compounds which produce known metabolic and toxicological responses.

Benzo(a)pyrene is a known rodent and likely human carcinogen and is the prototype of a class of compounds, the polycyclic aromatic hydrocarbons (PAH). It is metabolized by several forms of cytochrome P450 (P450 isozymes) and associated enzymes to form both activated and detoxified metabolites. The ultimate metabolites are the bay-region diol epoxide, benzo(a)pyrene-7,8-diol-9,10-epoxide (BPDE) and the K-region diol epoxide, 9-hydroxy benzo(a)pyrene-4,5-oxide, both of which induce formation of DNA adducts. DNA adducts have been shown to persist in rat liver up to 56 days following treatment with benzo(a)pyrene at a dose of 10 mg/kg body weight three times per week for two weeks (Qu and Stacey (1996) Carcinogenesis 17:53-59).

Acetaminophen is a widely-used analgesic. It is metabolized by specific cytochrome P450 isozymes with the majority of the drug undergoing detoxification by glucuronic acid, sulfate and glutathione conjugation pathways. However, at supratherapeutic doses, acetaminophen is metabolized to an active intermediate, *N*-acetyl-*p*-benzoquinone imine (NAPQI) which can cause hepatic and renal failure. NAPQI then binds to sulfhydryl groups of proteins causing their inactivation and leading to subsequent cell death (Kroger et al. (1997) Gen. Pharmacol. 28:257-263).

Clofibrate is an hypolidemic drug which lowers elevated levels of serum triglycerides. In rodents, chronic treatment produces hepatomegaly and an increase in hepatic peroxisomes (peroxisome

proliferation). Peroxisome proliferators (PPs) are a class of drugs which activate the PP-activated receptor in rodent liver, leading to enzyme induction, stimulation of S-phase, and a suppression of apoptosis (Hasmall and Roberts (1999) Pharmacol. Ther. 82:63-70). PPs include the fibrate class of hypolidemic drugs, phenobarbitone, thiazolidinediones, certain non-steroidal anti-inflammatory drugs, and naturally-occuring fatty acid-derived molecules (Gelman et al. (1999) Cell. Mol. Life Sci. 55:932-943). Clofibrate has been shown to increase levels of cytochrome P450 4A. It is also involved in transcription of β-oxidation genes as well as induction of PP-activated receptors (Kawashima et al. (1997) Arch. Biochem. Biophys. 347:148-154). Peroxisome proliferation that is induced by both clofibrate and the chemically-related compound fenofibrate is mediated by a common inhibitory effect on mitochondrial membrane depolarization (Zhou and Wallace (1999) Toxicol. Sci. 48:82-89).

Toxicological effects in the liver are also induced by other compounds. These can include carbon tetrachloride (a necrotic agent), hydrazine (a steatotic agent), α-naphthylisothiocyanate (a cholestatic agent), 4-acetylaminofluorene (a liver mitogen), and their corresponding metabolites, which are used in experimental protocols to measure toxicological responses (Waterfield et al. (1993) Arch. Toxicol. 67:244-254).

The present invention provides mammalian nucleic acid and protein molecules, their use in diagnostic and therapeutic applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

DISCLOSURE OF INVENTION

The invention provides a method for detecting or diagnosing the effect of a test compound or molecule associated with increased or decreased levels of nucleic acid molecules in a mammalian subject. The method comprises treating a mammalian subject with a known toxic compound or molecule which elicits a toxicological response, measuring levels of a plurality of nucleic acid molecules, selecting from the plurality of nucleic acid molecules those nucleic acid molecules that have levels modulated in samples treated with known toxic compounds or molecules when compared with untreated samples. Some of the levels may be upregulated by a toxic compound or molecule, others may be downregulated by a toxic compound or molecule, and still others may be upregulated with one known toxic compound or molecule and be downregulated with another known toxic compound or molecule. The selected nucleic acid molecules which are upregulated and downregulated by a known toxic compound or molecule are arrayed upon a substrate. The method further comprises measuring levels of nucleic acid molecules in the sample after the sample is treated with the toxic compound or molecule. Levels of nucleic acid molecules in a sample so treated are then compared with the plurality of the arrayed nucleic acid molecules to identify which sample nucleic acid molecules are upregulated and downregulated by the test

compound or molecule. In one embodiment, the nucleic acid molecules are hybridizable array elements of a microarray.

Preferably, the comparing comprises contacting the arrayed nucleic acid molecules with the sample nucleic acid molecules under conditions effective to form hybridization complexes between the arrayed nucleic acid molecules and the sample nucleic acid molecules; and detecting the presence or absence of the hybridization complexes. In this context, similarity may mean that at least 1, preferably at least 5, more preferably at least 10, of the upregulated arrayed nucleic acid molecules form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a greater extent than would the probes derived from a sample not treated with the test compound or molecule or a known toxic compound or molecule. Similarity may also mean that at least 1, preferably at least 5, more preferably at least 10, of the downregulated arrayed nucleic acid molecules form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a lesser extent than would the sample nucleic acid molecules of a sample not treated with the test compound or a known toxic compound. In one aspect, the arrayed nucleic acid molecules comprise SEQ ID NOs:1-47 or fragments thereof.

Preferred toxic compounds are selected from the group consisting of hypolipidemic drugs, n-alkylcarboxylic acids, n-alkylcarboxylic acid precursors, azole antifungal compounds, leukotriene D4 antagonists, herbicides, pesticides, phthalate esters, phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol and their corresponding metabolites, acetaminophen and its corresponding metabolites, benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like, carbon tetrachloride, hydrazine, α-naphthylisothiocyanate, 4-acetylaminofluorene, and their corresponding metabolites. Preferred tissues are selected from the group consisting of liver, kidney, brain, spleen, pancreas and lung.

The arrayed nucleic acid molecules comprise fragments of messenger RNA transcripts of genes that are upregulated-or-downregulated at least 2-fold, preferably at least 2.5-fold, more preferably at least 3-fold, in tissues treated with known toxic compounds when compared with untreated tissues. Preferred arrayed nucleic acid molecules are selected from the group consisting of SEQ ID NOs:1-47 or fragments thereof, some of whose expression is upregulated following treatment with a toxic compound or molecule and others of whose expression is downregulated following treatment with a toxic compound or molecule. More preferable are SEQ ID NOs:2, 4, 6, 8, 9, and 11 which are upregulated following treatment with a toxic compound or molecule, and SEQ ID NOs:1, 4, and 7 which are downregulated following treatment with a toxic compound or molecule.

The invention also provides a method comprising measuring levels of nucleic acid molecules in a sample after the sample is treated with a test compound or molecule. Levels of nucleic acid molecules in

a sample so treated are then compared with the plurality of the arrayed nucleic acid molecules to identify which sample nucleic acid molecules are upregulated and downregulated by the test compound or molecule. In one embodiment, the nucleic acid molecules are hybridizable array elements of a microarray.

Alternatively, the invention provides methods for screening a sample for a metabolic response to a test compound or molecule.

Alternatively, the invention provides methods for screening a test compound or molecule for a previously unknown metabolic response.

In another aspect, the invention provides methods for preventing a toxicological response by administering complementary nucleotide molecules against one or more selected upregulated nucleic acid molecules or a ribozyme that specifically cleaves such molecules. Alternatively, a toxicological response may be prevented by administering sense nucleotide molecules for one or more selected downregulated nucleic acid molecules.

In yet another aspect, the invention provides methods for preventing a toxicological response by administering an agonist which initiates transcription of a gene comprising a downregulated nucleic acid molecule of the invention. Alternatively, a toxicological response may be prevented by administering an antagonist which prevents transcription of a gene comprising an upregulated nucleic acid molecule of the invention.

In another aspect, the invention provides nucleic acid molecules whose transcript levels are modulated in a sample during a metabolic response to a toxic compound or molecule. The invention also provides nucleic acid molecules whose transcript levels are upregulated in a sample during a metabolic response to a toxic compound or molecule. The invention also provides nucleic acid molecules whose transcript levels are downregulated in a sample during a metabolic response to a toxic compound or molecule. Upregulation or downregulation is at least 2-fold, more preferably at least 2.5-fold, even more preferably at least 3-fold. The metabolic response to a toxic compound or molecule may be a toxicological response. The invention also provides mammalian nucleic acid molecules which are homologous to the upregulated and downregulated nucleic acid molecules. In one aspect, preferred arrayed nucleic acid molecules are selected from the group consisting of SEQ ID NOs:1-47, or fragments thereof.

The invention also provides a method for using a molecule selected from SEQ ID NOs:1-59 or a portion thereof to screen a library of molecules to identify at least one ligand which specifically binds the selected molecule, the method comprising combining the selected molecule with the library of molecules under conditions allowing specific binding, and detecting specific binding, thereby identifying a ligand which specifically binds the selected molecule.

Such libraries include DNA and RNA molecules, peptides, peptide nucleic acids, agonists, antagonists, antibodies, immunoglobulins, drug compounds, pharmaceutical agents, and other ligands. In one aspect, the ligand identified using the method modulates the activity of the selected molecule. In an analogous method, the selected molecule or a portion thereof is used to purify a ligand. The method involves combining the selected molecule or a portion thereof with a sample under conditions to allow specific binding, detecting specific binding between the selected molecule and ligand, recovering the bound selected molecule, and separating the selected molecule from the ligand to obtain purified ligand. The invention further provides a method for using at least a portion of the proteins encoded by SEQ ID NOs:1-47 and the proteins of SEQ ID NOs:48-59 to produce antibodies.

The invention further provides a method for inserting a marker gene into the genomic DNA of an animal to disrupt the expression of the natural nucleic acid molecule. The invention also provides a method for using the nucleic acid molecule to produce an animal model system, the method comprising constructing a vector containing the nucleic acid molecule; introducing the vector into a totipotent embryonic stem cell; selecting an embryonic stem cell with the vector integrated into genomic DNA; microinjecting the selected cell into a blastocyst, thereby forming a chimeric blastocyst; transferring the chimeric blastocyst into a pseudopregnant dam, wherein the dam gives birth to a chimeric animal containing at least one additional copy of nucleic acid molecule in its germ line; and breeding the chimeric animal to generate a homozygous animal model system.

The invention also provides a substantially purified mammalian protein or a portion thereof. The invention further provides isolated and purified proteins encoded by the nucleic acid molecules of SEQ ID NOs:1-11, 17-33, 36, 39, and 41. The invention further provides isolated and purified protein molecule of SEQ ID NOs:50 and 53. Additionally, the invention provides a pharmaceutical composition comprising a substantially purified mammalian protein or a portion thereof in conjunction with a pharmaceutical carrier.

The invention further provides an isolated and purified mammalian nucleic acid molecule variant having at least 70% nucleic acid sequence identity to the mammalian nucleic acid molecule selected from SEQ ID NO:1-47 and fragments thereof. The invention also provides an isolated and purified nucleic acid molecule having a sequence which is complementary to the mammalian nucleic acid molecule comprising a nucleic acid molecule selected from SEQ ID NO:1-47 and fragments thereof.

The invention further provides an expression vector containing at least a fragment of the mammalian nucleic acid molecule selected from the group consisting of SEQ ID NOs:1-47. In another aspect, the expression vector is contained within a host cell.

The invention also provides a method for producing a mammalian protein, the method comprising the steps of: (a) culturing the host cell containing an expression vector containing a

mammalian nucleic acid molecule of the invention under conditions suitable for the expression of the polypeptide; and (b) recovering the polypeptide from the host cell culture.

The invention also provides a pharmaceutical composition comprising a substantially purified mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and the amino acid sequence of SEQ ID NOs:50 and 53 and fragments thereof, in conjunction with a suitable pharmaceutical carrier.

The invention further includes an isolated and purified antibody which binds to a mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOs:50 and 53 or fragments thereof. The invention also provides a purified agonist and a purified antagonist.

BRIEF DESCRIPTION OF THE SEQUENCE LISTING

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The Sequence Listing contains the nucleic acid sequence of exemplary mammalian nucleic acid molecules of the invention, SEQ ID NOs:1-47, 60-135, 137, and 138; the protein sequence of exemplary mammalian protein molecules of the invention, SEQ ID NOs:48-59 and 136.

MODES FOR CARRYING OUT THE INVENTION

Definitions

"Sample" is used in its broadest sense. A sample containing nucleic acid molecules may comprise a bodily fluid; a cell; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; genomic DNA, RNA, or cDNA in solution or bound to a substrate; a biological tissue or biopsy thereof; a fingerprint or tissue print; natural or synthetic fibres; in a solution; in a liquid suspension; in a gaseous suspension; in an aerosol; and the like.

"Plurality" refers preferably to a group of one or more members, preferably to a group of at least about 10, and more preferably to a group of at least about 100 members, and even more preferably a group of 10,000 members.

"Substrate" refers to a rigid or semi-rigid support to which nucleic acid molecules or proteins are bound and includes membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, capillaries or other tubing, plates, polymers, and microparticles with a variety of surface forms including wells, trenches, pins, channels and pores.

"Modulates" refers to a change in activity (biological, chemical, or immunological) or lifespan resulting from specific binding between a molecule and either a nucleic acid molecule or a protein.

"Microarray" refers to an ordered arrangement of hybridizable array elements on a substrate. The array elements are arranged so that there are preferably at least ten or more different array elements, more preferably at least 100 array elements, even more preferably at least 1000 array elements, and most preferably 10,000. Furthermore, the hybridization signal from each of the array elements is individually distinguishable. In a preferred embodiment, the array elements comprise nucleic acid molecules.

"Nucleic acid molecule" refers to a nucleic acid, oligonucleotide, nucleotide, polynucleotide or any fragment thereof. It may be DNA or RNA of genomic or synthetic origin, double-stranded or single-stranded, and combined with carbohydrate, lipids, protein, or other materials to perform a particular activity such as transformation or form a useful composition such as a peptide nucleic acid (PNA). "Oligonucleotide" is substantially equivalent to the terms amplimer, primer, oligomer, element, target, and probe and is preferably single stranded.

"Protein" refers to an amino acid sequence, oligopeptide, peptide, polypeptide, or portions thereof whether naturally occurring or synthetic. Exemplary portions are the first twenty consecutive amino acids of a mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOs:50 and 53.

"Up-regulated" refers to a nucleic acid molecule whose levels increased in a treated sample compared with the nucleic acid molecule in an untreated sample.

"Down-regulated" refers to nucleic acid molecule whose levels decreased in a treated sample compared with the nucleic acid molecule in an untreated sample.

"Toxic compound" or "toxic agent" is any compound, molecule, or agent that elicits a biochemical, metabolic, and physiological response in an individual or animal, such as i) DNA damage, ii) cell damage, iii) organ damage or cell death, or iv) clinical morbidity or mortality.

"Toxicological response" refers to a biochemical, metabolic, and physiological response in an individual or animal which has been exposed to a toxic compound or agent.

"Fragment" refers to an Incyte clone or any part of a molecule which retains a usable, functional characteristic. Useful fragments include oligonucleotides and polynucleotides which may be used in hybridization or amplification technologies or in regulation of replication, transcription or translation. Exemplary fragments are the first sixty consecutive nucleotides of SEQ ID NOs:1-47. Useful fragments also include polypeptides and protein molecules which have antigenic potential and which may be used with a suitable pharmaceutical carrier in a pharmaceutical composition. Exemplary fragments are the first twenty consecutive amino acids of a mammalian protein encoded by SEQ ID NOs:1-11, 17-33, 36, 39, and 41 and mammalian protein of SEQ ID NOs:50 and 53.

"Hybridization complex" refers to a complex between two nucleic acid molecules by virtue of the formation of hydrogen bonds between purines and pyrimidines.

"Ligand" refers to any compound, molecule, or agent which will bind specifically to a complementary site on a nucleic acid molecule or protein. Such ligands stabilize or modulate the activity of nucleic acid molecules or proteins of the invention and may be composed of at least one of the following: inorganic and organic substances including nucleic acids, proteins, carbohydrates, fats, and lipids.

"Percent identity" or "% identity" refers to the percentage of sequence similarity found in a comparison of two or more amino acid or nucleic acid sequences. Percent identity can be determined electronically, e.g., by using the MEGALIGN program (DNASTAR, Madison WI) which creates alignments between two or more sequences according to methods selected by the user, e.g., the clustal method. (See, e.g., Higgins, D.G. and P.M. Sharp (1988) Gene 73:237-244.) The clustal algorithm groups sequences into clusters by examining the distances between all pairs. The clusters are aligned pairwise and then in groups. The percentage similarity between two amino acid sequences, e.g., sequence A and sequence B, is calculated by dividing the length of sequence A, minus the number of gap residues in sequence A, minus the number of gap residues in sequence B, into the sum of the residue matches between sequence A and sequence B, times one hundred. Gaps of low or of no similarity between the two amino acid sequences are not included in determining percentage similarity. Percent identity between nucleic acid sequences can also be counted or calculated by other methods known in the art, e.g., the Jotun Hein method. (See, e.g., Hein, J. (1990) Methods Enzymol. 183:626-645.) Identity between sequences can also be determined by other methods known in the art, e.g., by varying hybridization conditions.

"Substantially purified" refers to nucleic acid molecules or proteins that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably about 75% free, and most preferably about 90% free, from other components with which they are naturally associated.

The Invention

The present invention provides mammalian nucleic acid and protein molecules and method of using the nucleic acid molecules for screening test compounds and molecules for toxicological responses. Additionally the invention provides methods for characterizing the toxicological responses of a sample to a test compound or molecule. In particular, the present invention provides a composition comprising a plurality of nucleic acid molecules derived from human cDNA libraries, monkey cDNA libraries, mouse cDNA libraries, normal rat liver cDNA libraries, normalized rat liver cDNA libraries, prehybridized rat liver cDNA libraries, subtracted rat liver cDNA libraries, and rat kidney cDNA libraries. The nucleic acid molecules have been further selected for exhibiting upregulated or downregulated gene expression

in rat livers when the rats have been exposed to a known hepatotoxin, including a peroxisomal proliferator (PP), acetaminophen or one of its corresponding metabolites, a polycyclic aromatic hydrocarbon (PAH), carbon tetrachloride, hydrazine, α-naphthylisothiocyanate, 4-acetylaminofluorene, and their corresponding metabolites.

PPs include hypolipidemic drugs, such as clofibrate, fenofibrate, clofenic acid, nafenopin, gemfibrozil, ciprofibrate, bezafibrate, halofenate, simfibrate, benzofibrate, etofibrate, WY-14,643, and the like; n-alkylcarboxylic acids, such as trichloroacetic acid, valproic acid, hexanoic acid, and the like; n-alkylcarboxylic acid precursors, such as trichloroethylene, etrachloroethylene, and the like; azole antifungal compounds, such as bifonazole, and the like; leukotriene D4 antagonists; herbicides; pesticides; phthalate esters, such as di-[2-ethylhexyl] phthalate, mono-[2-ethylhexyl] phthalate, and the like; and natural chemicals, such as phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol, and the like. In a preferred embodiment the toxin is clofibrate, or one of its corresponding metabolites. In another prefered embodiment the toxin is fenofibrate, or one of its corresponding metabolites.

PAHs include compounds such as benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like. In a preferred embodiment the toxin is benzo(a)pyrene, or one of its corresponding metabolites.

SEQ ID NOs:1-16 were identified by their pattern of at least two-fold upregulation or downregulation following hybridization with sample nucleic acid molecules from rat liver tissue treated with a known toxic compound. SEQ ID NOs:17-47 were identified by their homology to the sample nucleic acid molecules from rat liver tissue treated with a known toxic compound. These and other nucleic acid molecules can be immobilized on a substrate as hybridizable array elements in a microarray format. The microarray may be used to characterize gene expression patterns associated with novel compounds to elucidate any toxicological responses or to monitor the effects of treatments during clinical trials or therapy where metabolic responses to toxic compounds may be expected.

When the nucleic acid molecules are employed as hybridizable array elements in a microarray, the array elements are organized in an ordered fashion so that each element is present at a specified location on the substrate. Because the array elements are at specified locations on the substrate, the hybridization patterns and intensities (which together create a unique expression profile) can be interpreted in terms of expression levels of particular genes and can be correlated with a toxicological response associated with a test compound or molecule.

The invention also provides a substantially purified and isolated mammalian protein comprising the protein molecule of SEQ ID NOs:50 and 53 or portion thereof. The invention further provides isolated and purified proteins encoded by the nucleic acid molecules of SEQ ID NOs:1-11, 17-33, 36, 39,

and 41, or portion thereof.

Furthermore, the present invention provides methods for screening test compounds or therapeutics for potential toxicological responses and for screening a sample's toxicological response to a particular test compound or molecule. Briefly, these methods entail treating a sample with the test compound or molecule to elicit a change in gene expression patterns comprising the expression of a plurality of sample nucleic acid molecules. Nucleic acid molecules are selected by identifying those genes in rat liver or kidney that are upregulated-or-downregulated at least 2-fold, more preferably at least 2.5-fold, most preferably at least 3-fold, when treated with a known toxic compound or molecule. The nucleic acid molecules are arrayed on a substrate. Then, the arrayed nucleic acid molecules and sample nucleic acid molecules are combined under conditions effective to form hybridization complexes which may be detected by methods well known in the art. Detection of higher or lower levels of such hybridization complexes compared with hybridization complexes derived from untreated samples and samples treated with a compound that is known not to induce a toxicological response correlates with a toxicological response of a test compound or a toxicological response to a molecule.

Complementary DNA libraries

Molecules are identified that reflect all or most of the genes that are expressed in rat liver or kidney. Molecules may be identified by isolating clones derived from several types of rat cDNA libraries, including normal rat cDNA libraries, normalized rat cDNA libraries, prehybridized rat cDNA libraries, and subtracted cDNA libraries. Clone inserts derived from these clones may be partially sequenced to generate expressed sequence tags (ESTs). Molecules are also identified by comparing the clones from rat cDNA libraries with clones from human, monkey, and mouse cDNA libraries using computer software nucleic acid comparison programs such as BLAST (see, e.g., Altschul, S.F. (1993) J. Mol. Evol. 3:290-300; Altschul, et al. (1990) J. Mol. Biol. 215:403-410).

In one embodiment, two collections of ESTs are identified and sequenced. A first collection of ESTs (the originator molecules) are derived from rat liver and kidney and are derived from the cDNA libraries presented in the Examples. A second collection includes ESTs derived from other rat cDNA libraries available in the ZOOSEQ database (Incyte Pharmaceuticals, Inc. Palo Alto CA).

The two collections of ESTs are clustered electronically to form master clusters of ESTs. Master clusters are formed by identifying overlapping EST molecules and assembling these ESTs. A nucleic acid fragment assembly tool, such as the Phrap tool (Phil Green, University of Washington) and the GELVIEW fragment assembly system (GCG, Madison WI), can be used for this purpose. The minimum number of clones which constitute a cluster is two. In another embodiment, a collection of human genes known to be expressed in response to toxic agents are used to select representative ESTs from the 113 rat cDNA libraries. The master cluster process is repeated for these molecules.

After assembling the clustered consensus nucleic acid sequences, a representative 5' clone is nominated from each master cluster. The most 5' clone is preferred because it is most likely to contain the complete gene. The nomination process is described in greater detail in "Relational Database and System for Storing Information Relating to Biomolecular Sequences and Reagents", USSN 09/034,807, filed March 4, 1998, herein incorporated in its entirety by reference. The EST molecules are used as array elements on a microarray.

Selection of arrayed nucleic acid molecules

Samples are treated, preferably at subchronic doses, with one or more known toxic compounds over a defined time course. Preferably, the agents are peroxisomal proliferators (PPs), acetaminophen or one of its corresponding metabolites, polycyclic aromatic hydrocarbons (PAHs), carbon tetrachloride, hydrazine, α-naphthylisothiocyanate, 4-acetylaminofluorene, or their corresponding metabolites.

The gene expression patterns derived from such treated biological samples can be compared with the gene expression patterns derived from untreated biological samples to identify and select nucleic acid molecules whose expression is either upregulated or downregulated due to the response to the toxic compounds. These selected molecules may then be employed as array elements alone or in combination with other array element molecules. Such a microarray is particularly useful to detect and characterize gene expression patterns associated with known toxic compounds. Such gene expression patterns can then be used for comparison to identify other compounds which also elicit a toxicological response.

The arrayed nucleic acid molecules can be manipulated to optimize their performance in hybridization. To optimize hybridization, the arrayed nucleic acid molecules are examined using a computer algorithm to identify portions of genes without potential secondary structure. Such computer algorithms are well known in the art and are part of OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or LASERGENE software (DNASTAR, Madison WI). These programs can search within nucleic acid sequences to identify stem loop structures and tandem repeats and to analyze G + C content of the sequence (those molecules with a G + C content greater than 60% are excluded). Alternatively, the arrayed nucleic acid molecules can be optimized by trial and error. Experiments can be performed to determine whether sample nucleic acid molecules and complementary arrayed nucleic acid molecules hybridize optimally under experimental conditions.

The arrayed nucleic acid molecules can be any RNA-like or DNA-like material, such as mRNAs, cDNAs, genomic DNA, peptide nucleic acids, branched DNAs and the like. The arrayed nucleic acid molecules can be in sense or antisense orientations.

In one embodiment, the arrayed nucleic acid molecules are cDNAs. The size of the DNA sequence of interest may vary, and is preferably from 50 to 10,000 nucleotides, more preferably from 150 to 3,500 nucleotides. In a second embodiment, the nucleic acid molecules are vector DNAs. In this case

the size of the DNA sequence of interest, i.e., the insert sequence, may vary from about 50 to 10,000 nucleotides, more preferably from about 150 to 3,500 nucleotides.

The nucleic acid molecule sequences of the Sequence Listing have been prepared by current, state-of-the-art, automated methods and, as such, may contain occasional sequencing errors and unidentified nucleotides. Nucleotide analogues can be incorporated into the nucleic acid molecules by methods well known in the art. The only requirement is that the incorporated nucleotide analogues must serve to base pair with sample nucleic acid molecules. For example, certain guanine nucleotides can be substituted with hypoxanthine which base pairs with cytosine residues. However, these base pairs are less stable than those between guanine and cytosine. Alternatively, adenine nucleotides can be substituted with 2,6-diaminopurine which can form stronger base pairs than those between adenine and thymidine. Additionally, the nucleic acid molecules can include nucleotides that have been derivatized chemically or enzymatically. Typical modifications include derivatization with acyl, alkyl, aryl or amino groups.

The nucleic acid molecules can be immobilized on a substrate via chemical bonding. Furthermore, the molecules do not have to be directly bound to the substrate, but rather can be bound to the substrate through a linker group. The linker groups are typically about 6 to 50 atoms long to provide exposure to the bound nucleic acid molecule. Preferred linker groups include ethylene glycol oligomers, diamines, diacids and the like. Reactive groups on the substrate surface react with one of the terminal portions of the linker to bind the linker to the substrate. The other terminal portion of the linker is then functionalized for binding the nucleic acid molecule. Preferred substrates are any suitable rigid or semirigid support, including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which the arrayed nucleic acid molecules are bound.

The samples can be any sample comprising sample nucleic acid molecules and obtained from any bodily fluid (blood, urine, saliva, phlegm, gastric juices, etc.), cultured cells, biopsies, or other tissue preparations. The samples can be derived from any species, but preferably from eukaryotic species, and more preferably from mammalian species such as rat and human.

DNA or RNA can be isolated from the sample according to any of a number of methods well known to those of skill in the art. For example, methods of purification of nucleic acids are described in Tijssen, P. (1993) Laboratory Techniques in Biochemistry and Molecular Biology: Hybridization With Nucleic Acid Probes, Part I. Theory and Nucleic Acid Preparation, Elsevier, New York, NY. In one preferred embodiment, total RNA is isolated using the TRIZOL total RNA isolation reagent (Life Technologies, Inc., Gaithersburg MD) and mRNA is isolated using oligo d(T) column chromatography or

glass beads. When sample nucleic acid molecules are amplified it is desirable to amplify the sample nucleic acid molecules and maintain the relative abundances of the original sample, including low abundance transcripts. RNA can be amplified in vitro, in situ, or in vivo (See Eberwine US Patent No. 5,514,545).

It is also advantageous to include controls within the sample to assure that amplification and labeling procedures do not change the true distribution of nucleic acid molecules in a sample. For this purpose, a sample is spiked with an amount of a control nucleic acid molecule predetermined to be detectable upon hybridization to its complementary arrayed nucleic acid molecule and the composition of nucleic acid molecules includes reference nucleic acid molecules which specifically hybridize with the control arrayed nucleic acid molecules. After hybridization and processing, the hybridization signals obtained should reflect accurately the amounts of control arrayed nucleic acid molecules added to the sample.

Prior to hybridization, it may be desirable to fragment the sample nucleic acid molecules. Fragmentation improves hybridization by minimizing secondary structure and cross-hybridization to other sample nucleic acid molecules in the sample or noncomplementary nucleic acid molecules. Fragmentation can be performed by mechanical or chemical means.

Labeling

The sample nucleic acid molecules may be labeled with one or more labeling moieties to allow for detection of hybridized arrayed/sample nucleic acid molecule complexes. The labeling moieties can include compositions that can be detected by spectroscopic, photochemical, biochemical, bioelectronic, immunochemical, electrical, optical or chemical means. The labeling moieties include radioisotopes, such as ^{32}P, ^{33}P or ^{35}S, chemiluminescent compounds, labeled binding proteins, heavy metal atoms, spectroscopic markers, such as fluorescent markers and dyes, magnetic labels, linked enzymes, mass spectrometry tags, spin labels, electron transfer donors and acceptors, and the like. Preferred fluorescent markers include Cy3 and Cy5 fluorophores (Amersham Pharmacia Biotech, Piscataway NJ).

Hybridization

The nulceic acid molecule sequence of SEQ ID NOs:1-47 and fragments thereof can be used in various hybridization technologies for various purposes. Hybridization probes may be designed or derived from SEQ ID NOs:1-47. Such probes may be made from a highly specific region such as the 5' regulatory region or from a conserved motif, and used in protocols to identify naturally occurring sequences encoding the mammalian protein, allelic variants, or related sequences, and should preferably have at least 50% sequence identity to any of the protein sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NOs:1-47 or from genomic sequences including promoters, enhancers, and introns of the mammalian gene.

Hybridization or PCR probes may be produced using oligolabeling, nick translation, end-labeling, or PCR amplification in the presence of the labeled nucleotide. A vector containing the nucleic acid sequence may be used to produce an mRNA probe in vitro by addition of an RNA polymerase and labeled nucleic acid molecules. These procedures may be conducted using commercially available kits such as those provided by Amersham Pharmacia Biotech.

The stringency of hybridization is determined by G+C content of the probe, salt concentration, and temperature. In particular, stringency can be increased by reducing the concentration of salt or raising the hybridization temperature. In solutions used for some membrane based hybridizations, additions of an organic solvent such as formamide allows the reaction to occur at a lower temperature. Hybridization can be performed at low stringency with buffers, such as 5 x SSC with 1% sodium dodecyl sulfate (SDS) at 60°C, which permits the formation of a hybridization complex between nucleotide sequences that contain some mismatches. Subsequent washes are performed at higher stringency with buffers such as 0.2 x SSC with 0.1% SDS at either 45°C (medium stringency) or 68°C (high stringency). At high stringency, hybridization complexes will remain stable only where the nucleic acid sequences are completely complementary. In some membrane-based hybridizations, preferably 35% or most preferably 50%, formamide can be added to the hybridization solution to reduce the temperature at which hybridization is performed, and background signals can be reduced by the use of other detergents such as Sarkosyl or Triton X-100 and a blocking agent such as salmon sperm DNA. Selection of components and conditions for hybridization are well known to those skilled in the art and are reviewed in Ausubel (supra) and Sambrook et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY.

Hybridization specificity can be evaluated by comparing the hybridization of specificity-control nucleic acid molecules to specificity-control sample nucleic acid molecules that are added to a sample in a known amount. The specificity-control arrayed nucleic acid molecules may have one or more sequence mismatches compared with the corresponding arrayed nucleic acid molecules. In this manner, whether only complementary arrayed nucleic acid molecules are hybridizing to the sample nucleic acid molecules or whether mismatched hybrid duplexes are forming is determined.

Hybridization reactions can be performed in absolute or differential hybridization formats. In the absolute hybridization format, nucleic acid molecules from one sample are hybridized to the molecules in a microarray format and signals detected after hybridization complex formation correlate to nucleic acid molecule levels in a sample. In the differential hybridization format, the differential expression of a set of genes in two biological samples is analyzed. For differential hybridization, nucleic acid molecules from both biological samples are prepared and labeled with different labeling moieties. A mixture of the two labeled nucleic acid molecules is added to a microarray. The microarray is then examined under

conditions in which the emissions from the two different labels are individually detectable. Molecules in the microarray that are hybridized to substantially equal numbers of nucleic acid molecules derived from both biological samples give a distinct combined fluorescence (Shalon et al. PCT publication WO95/35505). In a preferred embodiment, the labels are fluorescent markers with distinguishable emission spectra, such as Cy3 and Cy5 fluorophores.

After hybridization, the microarray is washed to remove nonhybridized nucleic acid molecules and complex formation between the hybridizable array elements and the nucleic acid molecules is detected. Methods for detecting complex formation are well known to those skilled in the art. In a preferred embodiment, the nucleic acid molecules are labeled with a fluorescent label and measurement of levels and patterns of fluorescence indicative of complex formation is accomplished by fluorescence microscopy, preferably confocal fluorescence microscopy.

In a differential hybridization experiment, nucleic acid molecules from two or more different biological samples are labeled with two or more different fluorescent labels with different emission wavelengths. Fluorescent signals are detected separately with different photomultipliers set to detect specific wavelengths. The relative abundances/expression levels of the nucleic acid molecules in two or more samples is obtained.

Typically, microarray fluorescence intensities can be normalized to take into account variations in hybridization intensities when more than one microarray is used under similar test conditions. In a preferred embodiment, individual arrayed-sample nucleic acid molecule complex hybridization intensities are normalized using the intensities derived from internal normalization controls contained on each microarray.

The labeled sample emits specific wavelengths which are detected using a plurality of photomultipliers. The nucleic acid molecules whose relative abundance/expression levels are modulated by treatment of a sample with a known toxic compound can be used as hybridizable elements in a microarray. Such a microarray can be employed to identify expression profiles associated with particular toxicological responses. Then, a particular subset of these photomultipliers set to detect specific wavelengths. The relative expression levels of the arrayed nucleic acid molecules can be identified as to which arrayed nucleic acid molecule expression is modulated in response to a particular toxicological agent. These photomultipliers are set to detect specific wavelengths. The relative expression levels of the nucleic acid molecules can be employed to identify other compounds with a similar toxicological response.

Alternatively, for some treatments with known side effects, the microarray, and expression patterns derived therefrom, is employed to prospectively define the treatment regimen. A dosage is established that minimizes expression patterns associated with undesirable side effects. This approach

may be more sensitive and rapid than waiting for the patient to show toxicological side effects before altering the course of treatment.

Generally, the method for screening a library of test compounds or molecules to identify those with a toxicological response entails selecting a plurality of arrayed genes whose expression levels are modulated in tissues treated with known toxic compounds when compared with untreated tissues. Then a sample is treated with the test compound or molecule to induce a pattern of gene expression comprising the expression of a plurality of sample nucleic acid molecules. Tissues from a mammalian subject treated at various dosages of the test compound may be screened to determine which doses may be toxic.

Then, the expression levels of the arrayed genes and the sample nucleic acid molecules are compared to identify those compounds that induce expression levels of the sample nucleic acid molecules that are similar to those of the arrayed genes. In one preferred embodiment, gene expression levels are compared by contacting the arrayed genes with the sample nucleic acid molecules under conditions effective to form hybridization complexes between arrayed genes and sample nucleic acid molecules; and detecting the presence or absence of the hybridization complexes.

Similarity may mean that at least 1, preferably at least 5, more preferably at least 10, of the upregulated arrayed genes form hybridization complexes with the sample nucleic acid molecules at least once during a time course to a greater extent than would the nucleic acid molecules of a sample not treated with the test compound. Similarity may also mean that at least 1, preferably at least 5, more preferably at least 10, of the downregulated nucleic acid molecules form hybridization complexes with the arrayed genes at least once during a time course to a lesser extent than would the nucleic acid molecules of a sample not treated with the test compound.

Such a similarity of expression patterns means that a toxicological response is associated with the compound or therapeutic tested. Preferably, the toxic compounds belong to the class of peroxisomal proliferators (PPs), including hypolipidemic drugs, such as clofibrate, fenofibrate, clofenic acid, nafenopin, gemfibrozil, ciprofibrate, bezafibrate, halofenate, simfibrate, benzofibrate, etofibrate, WY-14,643, and the like; n-alkylcarboxylic acids, such as trichloroacetic acid, valproic acid, hexanoic acid, and the like; n-alkylcarboxylic acid precursors, such as trichloroethylene, etrachloroethylene, and the like; azole antifungal compounds, such as bifonazole, and the like; leukotriene D4 antagonists; herbicides; pesticides; phthalate esters, such as di-[2-ethylhexyl] phthalate, mono-[2-ethylhexyl] phthalate, and the like; and natural chemicals, such as phenyl acetate, dehydroepiandrosterone (DHEA), oleic acid, methanol, and the like. In another embodiment, the toxic compound is acetaminophen or one of its corresponding metabolites. In yet another embodiment, the toxic compounds are polycyclic aromatic hydrocarbons (PAHs), including compounds such as benzo(a)pyrene, 3-methylcholanthrene, benz(a)anthracene, 7,12-dimethylbenz(a)anthracene, their corresponding metabolites, and the like. Of

particular interest is the study of the toxicological responses of these compounds on the liver, kidney, brain, spleen, pancreas, and lung.

Modification of Gene Expression Using Nucleic Acids

Gene expression may be modified by designing complementary or antisense molecules (DNA, RNA, or PNA) to the control, 5', 3', or other regulatory regions of the mammalian gene. Oligonucleotides designed with reference to the transcription initiation site are preferred. Similarly, inhibition can be achieved using triple helix base-pairing which inhibits the binding of polymerases, transcription factors, or regulatory molecules (Gee et al. In: Huber and Carr (1994) Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177). A complementary molecule may also be designed to block translation by preventing binding between ribosomes and mRNA. In one alternative, a library of nucleic acid molecules or fragments thereof may be screened to identify those which specifically bind a regulatory, nontranslated sequence .

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA followed by endonucleolytic cleavage at sites such as GUA, GUU, and GUC. Once such sites are identified, an oligonucleotide with the same sequence may be evaluated for secondary structural features which would render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing their hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary nucleic acids and ribozymes of the invention may be prepared via recombinant expression, in vitro or in vivo, or using solid phase phosphoramidite chemical synthesis. In addition, RNA molecules may be modified to increase intracellular stability and half-life by addition of flanking sequences at the 5' and/or 3' ends of the molecule or by the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. Modification is inherent in the production of PNAs and can be extended to other nucleic acid molecules. Either the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, and or the modification of adenine, cytidine, guanine, thymine, and uridine with acetyl-, methyl-, thio- groups renders the molecule less available to endogenous endonucleases.

Screening Assays

The nucleic acid molecule encoding the mammalian protein may be used to screen a library of molecules for specific binding affinity. The libraries may be DNA molecules, RNA molecules, PNAs, peptides, proteins such as transcription factors, enhancers, repressors, and other ligands which regulate the activity, replication, transcription, or translation of the nucleic acid molecule in the biological system. The assay involves combining the mammalian nucleic acid molecule or a fragment thereof with the

library of molecules under conditions allowing specific binding, and detecting specific binding to identify at least one molecule which specifically binds the nucleic acid molecule.

Similarly the mammalian protein or a portion thereof may be used to screen libraries of molecules in any of a variety of screening assays. The portion of the protein employed in such screening may be free in solution, affixed to an abiotic or biotic substrate (e.g. borne on a cell surface), or located intracellularly. Specific binding between the protein and molecule may be measured. Depending on the kind of library being screened, the assay may be used to identify DNA, RNA, or PNA molecules, agonists, antagonists, antibodies, immunoglobulins, inhibitors, peptides, proteins, drugs, or any other ligand, which specifically binds the protein. One method for high throughput screening using very small assay volumes and very small amounts of test compound is described in USPN 5,876,946, incorporated herein by reference, which screens large numbers of molecules for enzyme inhibition or receptor binding.

Purification of Ligand

The nucleic acid molecule or a fragment thereof may be used to purify a ligand from a sample. A method for using a mammalian nucleic acid molecule or a fragment thereof to purify a ligand would involve combining the nucleic acid molecule or a fragment thereof with a sample under conditions to allow specific binding, detecting specific binding, recovering the bound protein, and using an appropriate agent to separate the nucleic acid molecule from the purified ligand.

Similarly, the protein or a portion thereof may be used to purify a ligand from a sample. A method for using a mammalian protein or a portion thereof to purify a ligand would involve combining the protein or a portion thereof with a sample under conditions to allow specific binding, detecting specific binding between the protein and ligand, recovering the bound ligand, and using an appropriate chaotropic agent to separate the protein from the purified ligand.

Pharmacology

Pharmaceutical compositions are those substances wherein the active ingredients are contained in an effective amount to achieve a desired and intended purpose. The determination of an effective dose is well within the capability of those skilled in the art. For any compound, the therapeutically effective dose may be estimated initially either in cell culture assays or in animal models. The animal model is also used to achieve a desirable concentration range and route of administration. Such information may then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of protein or inhibitor which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity of such agents may be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., ED_{50} (the dose therapeutically effective in 50% of the population) and LD_{50} (the dose lethal to 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index, and it may be expressed as

the ratio, LD_{50}/ED_{50}. Pharmaceutical compositions which exhibit large therapeutic indexes are preferred. The data obtained from cell culture assays and animal studies are used in formulating a range of dosage for human use.

MODEL SYSTEMS

Animal models may be used as bioassays where they exhibit a toxic response similar to that of humans and where exposure conditions are relevant to human exposures. Mammals are the most common models, and most toxicity studies are performed on rodents such as rats or mice because of low cost, availability, and abundant reference toxicology. Inbred or outbred rodent strains provide a convenient model for investigation of the physiological consequences of under- or over-expression of genes of interest and for the development of methods for diagnosis and treatment of diseases. A mammal inbred to over-express a particular gene, so that the protein is secreted in milk, may also serve as a convenient source of the protein expressed by that gene.

Toxicology

Toxicology is the study of the effects of test compounds, molecules, or toxic agents on living systems to identify adverse effects. The majority of toxicity studies are performed on rats or mice to help predict whether adverse effects of agents will occur in humans. Observation of qualitative and quantitative changes in physiology, behavior, homeostatic, developmental, and reproductive processes, and lethality are used to generate profiles of safe or toxic responses and to assess the consequences on human health following exposure to the agent.

Toxicological tests measure the effects of a single, repeated, or long-term exposure of a subject to a substance. Substances may be tested for specific endpoints such as cytotoxicity, mutagenicity, carcinogenicity and teratogenicity. Degree of response varies according to the route of exposure (contact, ingestion, injection, or inhalation), age, sex, genetic makeup, and health status of the subject. Other tests establish the toxicokinetic and toxicodynamic properties of substances. Toxicokinetic studies trace the absorption, distribution in subject tissues, metabolism, storage, and excretion of substances. Toxicodynamic studies chart biological responses that are consequences of the presence of the substance in the subject tissues.

Genetic toxicology identifies and analyzes the ability of an agent to produce damage at a cellular or subcellular level. Such genotoxic agents usually have common chemical or physical properties that facilitate interaction with nucleic acids and are most harmful when mutated chromosomes are passed along to progeny. Toxicological studies may identify agents that increase the frequency of structural or functional abnormalities in progeny if administered to either parent before conception, to the mother during pregnancy, or to the developing organism. Mice and rats are most frequently used in these tests

because of their short reproductive cycle which allows investigators to breed sufficient quantities of individual animals to satisfy statistical requirements.

All types of toxicology studies on experimental animals involve preparation of a suitable form of the compound for administration, selection of the route of administration, and selection of a species which resembles the species of pharmacological interest. Dose concentrations of the compound are varied to identify, measure, and investigate a range of dose-related effects related to exposure.

Acute toxicity tests are based on a single administration of the agent to the subject to determine the symptomology or lethality of the agent. Three experiments are conducted; an experiment to define the initial dose range; an experiment to narrow the range of effective doses; and a final experiment to establish the dose-response curve.

Prolonged and subchronic toxicity tests are based on the repeated administration of the agent. Rat and dog are commonly used in these studies to provide data from species in different taxonomic orders. With the exception of carcinogenesis, there is considerable evidence that daily administration of an agent at high-dose concentrations for periods of three to four months will reveal most forms of toxicity in adult animals.

Chronic toxicity tests, with a duration of a year or more, are used to demonstrate either the absence of toxicity or the carcinogenic potential of an agent. When studies are conducted on rats, a minimum of at least one test group plus one control group are used. Animals are quarantined, examined for health, and monitored at the outset and at intervals throughout the experiment.

Transgenic Animal Models

Transgenic rodents which over-express or under-express a gene of interest may be inbred and used to model human diseases or to test therapeutic or toxic agents. (See USPN 4,736,866; USPN 5,175,383; and USPN 5,767,337; incorporated herein by reference). In some cases, the introduced gene may be activated at a specific time in a specific tissue type during fetal development or postnatally. Expression of the transgene is monitored by analysis of phenotype or tissue-specific mRNA expression, in transgenic animals before, during, and after being challenged with experimental drug therapies.

Embryonic Stem Cells

Embryonic stem cells (ES) isolated from rodent embryos retain the potential to form an embryo. When ES cells are placed inside a carrier embryo, they resume normal development and contribute to all tissues of the live-born animal. ES cells are the preferred cells used in the creation of experimental knockout and knockin rodent strains. Mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and are grown under culture conditions well known in the art. Vectors for knockout strains contain a disease gene candidate modified to include a marker gene which disrupts transcription and/or translation of the endogenous disease candidate gene in vivo. The vector is

introduced into ES cells by transformation methods such as electroporation, liposome delivery, microinjection, and the like which are well known in the art. The endogenous rodent gene is replaced by the disrupted disease gene through homologous recombination and integration during cell division. Expression of the marker gene confers a selective advantage to the transformed cells when incubated with an otherwise toxic/lethal selecting agent. Transformed ES cells are selected, identified, and preferably microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains.

ES cells are also used to study the differentiation of various cell types and tissues in vitro, such as neural cells, hematopoietic lineages, and cardiomyocytes (Bain et al. (1995) Dev. Biol. 168:342-357; Wiles and Keller (1991) Development 111:259-267; and Klug et al. (1996) J. Clin. Invest. 98:216-224). Recent developments demonstrate that ES cells derived from human blastocysts may also be manipulated in vitro to differentiate into eight separate cell lineages, including endoderm, mesoderm, and ectodermal cell types (Thomson et al. (1998) Science 282:1145-1147).

Knockout Analysis

In gene knockout analysis, a region of a human disease gene candidate is enzymatically modified to include a non-mammalian gene such as the neomycin phosphotransferase gene (*neo*; Capecchi (1989) Science 244:1288-1292). The inserted coding sequence disrupts transcription and translation of the targeted gene and prevents biochemical synthesis of the disease candidate protein. The modified gene is transformed into cultured embryonic stem cells (described above), the transformed cells are injected into rodent blastulae, and the blastulae are implanted into pseudopregnant dams. Transgenic progeny are crossbred to obtain homozygous inbred lines.

Knockin Analysis

Totipotent ES cells, present in the early stages of embryonic development, can be used to create knockin humanized animals (pigs) or transgenic animal models (mice or rats) of human diseases. With knockin technology, a region of a human gene is injected into animal ES cells, and the human sequence integrates into the animal cell genome by recombination. Totipotent ES cells which contain the integrated human gene are handled as described above. Inbred animals are studied and treated to obtain information on the analogous human condition. These methods have been used to model several human diseases. (See, e.g., Lee et al. (1998) Proc. Natl. Acad. Sci. 95:11371-11376; Baudoin et al. (1998) Genes Dev. 12:1202-1216; and Zhuang et al. (1998) Mol. Cell Biol. 18:3340-3349).

Non-Human Primate Model

The field of animal testing deals with data and methodology from basic sciences such as physiology, genetics, chemistry, pharmacology and statistics. These data are paramount in evaluating the

effects of therapeutic agents on non-human primates as they can be related to human health. Monkeys are used as human surrogates in vaccine and drug evaluations, and their responses are relevant to human exposures under similar conditions. Cynomolgus and Rhesus monkeys (Macaca fascicularis and Macaca mulatta, respectively) and Common Marmosets (Callithrix jacchus) are the most common non-human primates (NHPs) used in these investigations. Since great cost is associated with developing and maintaining a colony of NHPs, early research and toxicological studies are usually carried out in rodent models. In studies using behavioral measures such as drug addiction, NHPs are the first choice test animal. In addition, NHPs and individual humans exhibit differential sensitivities to many drugs and toxins and can be classified as a range of phenotypes from "extensive metabolizers" to "poor metabolizers" of these agents.

In additional embodiments, the nucleic acid molecules which encode the mammalian protein may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleic acid molecules that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Examples

It is understood that this invention is not limited to the particular methodology, protocols, and reagents described, as these may vary. It is also understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims. The examples below are provided to best describe the subject invention and its representative constituents.

I cDNA Library Construction

The RALINOT01 cDNA library was constructed from liver tissue removed from a pool of fifty 10- to 11-week-old Sprague-Dawley female rats (Pharmacon, Waverly PA). The animals were housed in standard laboratory caging and fed PMI-certified Rodent Diet #5002. The animals appeared to be in good health at the time tissue was harvested. The animals were anesthetized by CO_2 inhalation, and then cardiocentesis was performed.

Frozen tissue was homogenized and lysed in TRIZOL reagent (1 g tissue/10 ml TRIZOL; Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate, using a POLYTRON homogenizer (PT-3000; Brinkmann Instruments, Westbury NY). After a brief incubation on ice, chloroform (1:5 v/v) was mixed with the reagent, and then centrifuged at 1,000 rpm. The upper aqueous layer was removed to a fresh tube, and the RNA precipitated with isopropanol, resuspended in DEPC-treated water, and treated with DNase I for 25 min at 37°C. The RNA was re-extracted once with

phenol-chloroform, pH 4.7, and precipitated using 0.3 M sodium acetate and 2.5 volumes ethanol. The mRNA was then isolated using an OLIGOTEX kit (QIAGEN, Chatsworth CA) and used to construct the cDNA library.

The mRNA was handled according to the recommended protocols in the SUPERSCRIPT plasmid system (Life Technologies). The cDNAs were fractionated on a SEPHAROSE CL-4B column (Amersham Pharmacia Biotech), and those cDNAs exceeding 400 bp were ligated into the pINCY1 plasmid vector (Incyte Pharmaceuticals). The plasmid pINCY1 was subsequently transformed into DH5α or DH10B competent cells (Life Technologies).

The RAKINOT01 library was constructed using mRNA isolated from kidney tissue removed from a pool of fifty, 7- to 8-week-old male Sprague-Dawley rats, as described above.

The RAKINOT02 library was constructed using mRNA isolated from kidney tissue removed from a pool of fifty, 10- to 11-week-old female Sprague-Dawley rats, as described above.

II cDNA Library Normalization

In some cases, cDNA libraries were normalized in a single round according to the procedure of Soares et al. (1994, Proc. Natl. Acad. Sci. 91:9228-9232) with the following modifications. The primer to template ratio in the primer extension reaction was increased from 2:1 to 10:1. Reduction of each dNTP concentration in the reaction to 150μM allowed the generation of longer (400-1000 nucleotide (nt)) primer extension products. The reannealing hybridization was extended from 13 to 19 hours. The single stranded DNA circles of the normalized library were purified by hydroxyapatite chromatography, converted to partially double-stranded by random priming, and electroporated into DH10B competent bacteria (Life Technologies).

The Soares normalization procedure is designed to reduce the initial variation in individual cDNA frequencies and to achieve abundances within one order of magnitude while maintaining the overall sequence complexity of the library. In the normalization process, the prevalence of high-abundance cDNA clones decreases significantly, clones with mid-level abundance are relatively unaffected, and clones for rare transcripts are increased in abundance. In the modified Soares normalization procedure, significantly longer hybridization times are used to increase gene discovery rates by biasing the normalized libraries toward low-abundance cDNAs that are well represented in a standard transcript image.

The RALINON03, RALINON04, and RALINON07 normalized rat liver cDNA libraries were constructed with 2.0 x 10^6, 4.6 x 10^5, and 2.0 x 10^6 independent clones from the RALINOT01 cDNA library, respectively. The RALINOT01 cDNA library was normalized in one round using conditions adapted from Soares (supra) except that a significantly longer (48-hour) reannealing hybridization was

used.

III cDNA Library Prehybridization

The RALINOH01 cDNA library was constructed with clones from the RALINOT01 cDNA library. After preparation of the RALINOT01 cDNA library, 9,984 clones were spotted onto a nylon filter, lysed, and the plasmid DNA was bound to the filter. The filter was incubated with pre-warmed hybridization buffer and then hybridized at 42°C for 14-16 hours in 0.75 M NaCl, 0.1 M Na_2HPO_4/NaH_2PO_4, 0.15 M tris-HCl (pH 7.5), 5x Denhardt's Solution, 2% SDS, 100 µg/ml sheared salmon sperm DNA, 50% formamide, and [^{32}P]-labeled oligonucleotide molecules made from reverse transcribed rat liver mRNA from an untreated animal. The filter was rinsed with 2 x SSC (saline sodium citrate) at ambient temperature for 5 minutes followed by washing for 30 minutes at 68°C with pre-warmed washing solution (2 x SSC, 1% SDS). The wash was repeated with fresh washing solution for an additional 30 minutes at 68°C. Filters were then washed twice with pre-warmed washing solution (0.6 x SSC, 1% SDS) for 30 minutes at 68°C. Some 4,224 clones had very low hybridization signals and about 20% of the clones had no signals and two groups were isolated and sequenced.

IV Isolation and Sequencing of cDNA Clones

DNA was isolated using the following protocol. Single bacterial colonies were transferred into individual wells of 384-well plates (Genetix Ltd, Christchurch, United Kingdom) using sterile toothpicks. The wells contained 1 ml of sterile Terrific Broth (Life Technologies) with 25 mg/l carbenicillin and 0.4% glycerol (v/v). The plates were covered and placed in an incubator (Thermodyne, Newtown Square PA) at 37°C for 8-10 hours. Plasmid DNA was released from the cells and amplified using direct link PCR (Rao, V.B. (1994) Anal. Biochem. 216:1-14) as follows. The direct link PCR solution included 30 ml of NUCLEIX PLUS PCR nucleotide mix (Amersham Pharmacia Biotech, Piscataway NJ) and 300 µl of Taq DNA polymerase (Amersham Pharmacia Biotech). Five microlitres of the PCR solution were added to each of the 384 wells using the MICROLAB 2200 system (Hamilton, Reno NV); plates were centrifuged at 1000 rpm for 20 seconds and refrigerated until use. A 384 pin tool (V&P Scientific Inc, San Diego CA) was used to transfer bacterial cells from the incubation plate into the plate containing the PCR solution where 0.1% Tween 20 caused the cells to undergo lysis and release the plasmid DNA. After lysis, the plates were centrifuged up to 500 rpm, covered with a cycle sealer, and cycled using a 384-well DNA ENGINE thermal cycler (MJ Research, Watertown MA) using the program dPCR30 with the following parameters: Step 1) 95 °C, 1 minute; Step 2) 94 °C, 30 seconds; Step 3) 55 °C, 30 seconds; Step 4) 72 °C, 2 minutes; Step 5) steps 2, 3, and 4 repeated 29 times; Step 6) 72 °C, 10 minutes; and Step 7) storage at 4 °C.

The concentration of DNA in each well was determined by dispensing 100 µl PICO GREEN quantitation reagent (0.25% (v/v), Molecular Probes, Eugene OR) dissolved in 1x TE and 0.5 µl of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the quantitation reagent. The plate was scanned in a Fluoroscan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantitate the concentration of DNA. Typical concentrations of each DNA sample were in the range of 100 to 500 ng/ml.

The cDNAs were prepared for sequencing using either a HYDRA microdispenser (Robbins Scientific, Sunnyvale CA) or MICROLAB 2200 system (Hamilton) in combination with the DNA ENGINE thermal cyclers (MJ Research). The cDNAs were sequenced using the method of Sanger, F. and A.R. Coulson (J. Mol. Biol. (1975) 94:441-448) and the ABI 377 sequencing systems (PE Biosystems). Most of the isolates were sequenced according to standard ABI protocols using ABI kits (PE Biosystems). The solution volumes were used at 0.25x - 1.0x concentrations. Typically, 500 to 700 base pairs were sequenced in 3.5 to 4 hours. In the alternative, cDNAs may have been sequenced using solutions and dyes from Amersham Pharmacia Biotech.

V Rat Liver and Kidney Gene Selection

As a first step, originator molecules from high throughput sequencing experiments were derived from clone inserts from RALINOT01, RAKINOT01, RAKINOT02, RALINOH01, RALINON03, RALINON04 and RALINON07. cDNA library clones were obtained. There were 18,140 rat liver molecules and 5,779 rat kidney molecules.

Additionally, 1,500 rat molecules derived from clone inserts of any of 113 rat cDNA libraries were selected based on their homology to genes coding for polypeptides implicated in toxicological responses including peroxisome-associated genes, lysosome-associated genes, apoptosis-associated genes, cytochrome P450 genes, detoxification genes such as sulfotransferases, glutathione S-transferases, and cysteine proteases, and the like.

Then, all the remaining molecules derived from all of the rat cDNA library clones were clustered based on the originator molecules described above. The clustering process involved identifying overlapping molecules that have a match quality indicated by a product score of 50 using BLAST. 6581 master clusters were identified.

After forming the clone clusters, a consensus sequence was generated based on the assembly of the clone molecules using PHRAP (Phil Green, University of Washington). The assembled molecules were then annotated by first screening the assembled molecules against GenBank using BLASTn and then by screening the assembled molecules against GenPept using FASTX. About two thirds of the

assembled molecules were annotated, about one third of the assembled molecules were not annotated. For example, for nucleic acid sequence analysis, the program BLASTN 1.4.9MP-WashU was used with default parameters; ctxfactor=2.00; E=10; MatID, 0; Matrix name, +5,-4. In another example, for amino acid sequence analysis, the program NCBI-BLASTX 2.0.4 was used with default parameters; matrix, BLOSUM62; gap penalties, existence 11, extension 1; frameshift window, decay constant 50, 0.1.

VI Substrate and Array Element/Probe Preparation

Clones nominated in the process described in Example V were used to generate array elements. Each array element was amplified from bacterial cells. PCR amplification used primers complementary to the vector sequences flanking the cDNA insert. Array elements were amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μg. Amplified array elements were then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements were immobilized on polymer-coated glass slides. Glass microscope slides (Corning, Corning NY) cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides were etched in 4% hydrofluoric acid (VWR, West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma-Aldrich, St. Louis MO) in 95% ethanol. Coated slides were cured in a 110°C oven.

Array elements were applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522 and incorporated herein by reference. In brief, 1 μl of the array element DNA, at an average concentration of 0.5 μg/ml in 3 x SSC, was loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposited about 5 nl of the array element sample per slide. A total of 7404 array elements representing rat liver and kidney genes and a variety of control elements, including 14 synthetic control molecules, human genomic DNA, and yeast genomic DNA, were arrayed in four identical quadrants within a 1.8 cm^2 area of the glass substrate.

Microarrays were UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays were washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites were blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS; Tropix Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

VII Target Preparation

Male Sprague-Dawley rats (6-8 wk old) were dosed intraperitoneally with one of the following: clofibrate (CLO; Acros, Geel, Belgium) at 250 mg/kg body weight (bw); acetaminophen (APAP; Acros) at 1000 mg/kg bw; benzo(a)pyrene (B(a)P; Acros) at 10 mg/kg bw; or dimethylsulfoxide vehicle (DMSO;

Acros) at less than 2 ml/kg bw, and the animals were later euthanized by CO_2 inhalation. Animals were monitored daily for physical condition and body weight. Three animals per group were sacrificed approximately 12 hours, 24 hours, 3d (d), 7d, 14d, and 28d following the single dose. Prior to sacrifice a blood sample from each animal was drawn and assayed for serum alanine transferase (ALT) and serum aspartate aminotransferase (AST) levels using a diagnostic kit (Sigma-Aldrich). Observed gross pathology and liver weights were recorded at time of necropsy. Liver, kidney, brain, spleen and pancreas from each rat were harvested, flash frozen in liquid nitrogen, and stored at -80°C.

In the alternative, male Han-Wistar rats (8-9 wk old) were dosed by oral gavage with one of the following: fenofibrate (FEN; Sigma-Aldrich) at 250 mg/kg bw; carbon tetrachloride (CCL_4; Sigma-Aldrich) at 3160 mg/kg bw, hydrazine (HYDR; Sigma-Aldrich) at 120 mg/kg bw; α-naphthylisothiocyanate (ANIT; Sigma-Aldrich) at 200 mg/kg bw; 4-acetylaminofluorene (4-AFF; Lancaster Synthesis, Morecambe, Lancashire, UK) at 1000 mg/kg bw; corn oil vehicle, or sterile water vehicle, at 10 ml/kg bw. The animals were checked twice daily for clinical signs of distress. Blood was collected six days prior to the dose and at sacrifice. Three animals per group were sacrificed approximately six hours and 24 hours following the single dose. The animals were euthanized by exsanguination under isoflurane anaesthesia. Observed gross pathology and liver weights were recorded at time of necropsy. Livers from each rat were harvested, dissected into approximate 100 mg pieces, flash frozen in liquid nitrogen, and stored at -70°C.

For each target preparation, frozen liver was homogenized and lysed in TRIZOL reagent (Life Technologies, Gaithersburg MD) following the modifications for liver RNA isolation. Messenger RNA was isolated using an OLIGOTEX kit (QIAGEN) and labeled with either Cy3- or Cy5-labeled primers (Operon Technologies, Alameda CA) using the GEMBRIGHT labeling kit (Incyte Pharmaceuticals). Messenger RNA isolated from tissues of rats treated with clofibrate, acetaminophen, or benzo(a)pyrene was labeled with Cy5 and mRNA isolated from tissues of rats treated with DMSO was labeled with Cy3. Quantitative and differential expression pattern control cDNAs were added to each labeling reaction. Labeled cDNA was treated with 0.5 M sodium bicarbonate (pH 9.2) for 20 min at 85°C to degrade the RNA and purified using two successive CHROMA SPIN 30 gel filtration spin columns (Clontech, Palo Alto CA). Cy3-labeled control sample and Cy5-labeled experimental sample were combined and precipitated in glycogen, sodium acetate, and ethanol.

Targets are also prepared from tissue needle biopsy samples. Samples are used to identify changes within the tissue following exposure to, for example, a toxic compound, a potential toxic compound, a compound with unknown metabolic responses, and a pharmacological compound.

VIII Hybridization

Hybridizations were carried out using the methods described by Shalon (supra).

IX Detection

The microscope used to detect the reporter-labeled hybridization complexes was equipped with an Innova 70 mixed gas 10 W laser (Coherent Lasers, Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3, and 632 nm for excitation of Cy5. The excitation laser light was focused on the array using a 20x microscope objective (Nikon, Melville NY). The slide containing the array was placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example was scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excited the two fluorophores sequentially. Emitted light was split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics, San Jose CA) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes were used to filter the signals. The emission maxima of the fluorophores used were 565 nm for Cy3 and 650 nm for Cy5. Each array was typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus was capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans was typically calibrated using the signal intensity generated by a cDNA control species added to the probe mix at a known concentration. A specific location on the array contained a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two probes from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration was done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube was digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Norwood MA) installed in an IBM-compatible PC computer. The digitized data were displayed as an image where the signal intensity was mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data was also analyzed quantitatively. Where two different fluorophores were excited and measured simultaneously, the data were first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid was superimposed over the fluorescence signal image such that the signal from each spot

was centered in each element of the grid. The fluorescence signal within each element was then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis was the GEMTOOLS gene expression analysis program (Incyte Pharmaceuticals). .

X Results

The expression patterns of eight cytochrome P450 isozymes known to be induced in a toxicological response were monitored during the 28 day time course. The results using clofibrate, acetaminophen, and benzo(a)pyrene are shown in Table 1, Table 2, and Table 3, respectively. Each of the known genes was upregulated or downregulated greater than 2-fold at least once during the time course.

TABLE 1 Gene expression patterns (x-fold change) of known genes in clofibrate-treated rat liver

Gene	12 hours	24 hours	3 days	7 days	28 days
P450 LA-omega 4A3	14.8	26.6	1.1	0.5	0.47
P450 4A	7.0	16.6	1.4	0.5	1.3
P450 3A2	0.14	1.2	0.63	0.50	0.45

TABLE 2 Gene expression patterns (x-fold change) of known genes in acetaminophen-treated rat liver

Gene	12 hours	24 hours	3 days	7 days	14 days	28 days
P450 4A	1.0	4.5	2.1	2.0	4.4	4.8
P450f 2C7	0.21	0.43	0.47	0.5	1.2	1.3
P450 14DM	0.31	0.20	2.0	1.1	1.4	0.42

TABLE 3 Gene expression patterns (x-fold change) of known genes in benzo(a)pyrene-treated rat liver

Gene	12 hours	24 hours	3 days	7 days	14 days	28 days
P450 LA-omega 4A3	1.2	2.3	2.4	1.4	6.8	1.2
P450 MCA-inducible 1A2	7.3	9.2	5.7	2.5	2.5	0.5

In addition, results from two samples that had been treated identically were compared to determine the range of normal variation of gene expression patterns between the samples. In one analysis, where two different samples were prepared from identically treated tissues, gene expression patterns of cDNAs which were upregulated or downregulated not more than 1.7-fold were within the 95% confidence limits of a Poisson normal distribution. In a separate analysis, gene expression patterns of cDNAs which were upregulated or downregulated more than 2-fold accounted for not more than 5% of

the total hybridizable sample nucleic acid molecules in two identically-treated tissue samples.

We have discovered novel nucleotide molecules that are up-regulated or down-regulated at least 2-fold at least once during the time course. These molecules are SEQ ID NOs:1-16 provided in the Sequence Listing. These polynucleotide molecules can be used for screening compounds or therapeutics for a toxicologic effect and applications including detecting metabolic and toxicological responses, and in monitoring drug mechanism of action.

Table 4 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with clofibrate (CLO). Table 5 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with CLO.

TABLE 4 Gene expression patterns (x-fold change) of CLO-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	28 days
2	2.6	1.4	0.5	1.1	1.2
3	1.3	2	1.3	1.5	1.5
4	2	0.36	0.47	0.26	0.30
5	1.7	2.9	1.6	1.5	1.2
8	2.6	1.7	1.3	1.3	1.4

TABLE 5 Gene expression patterns (x-fold change) of CLO-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	28 days
1	n.d.	0.26	0.45	0.26	1.1
4	2.0	0.36	0.47	0.26	0.30
7	0.24	0.42	0.37	1.1	1.5

(n.d. = not detected)

Table 6 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with acetaminophen (APAP). Table 7 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with APAP.

TABLE 6 Gene expression patterns (x-fold change) of APAP-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	14 days	28 days
2	1.3	2.2	1.1	0.5	1.2	1.3
3	1.2	2.1	0.47	0.46	1.8	1.5
4	3.3	0.47	0.47	0.23	0.35	0.36
5	1.1	2.1	1.1	1.2	1.3	1.4
6	1.8	5	2.5	1.1	1.4	1.3
8	1.1	2.5	1.1	1	1.7	1.4

TABLE 7 Gene expression patterns (x-fold change) of APAP-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	24 hours	3 days	7 days	14 days	28 days
1	0.36	0.19	0.46	0.25	0.5	1.4
4	3.3	0.48	0.47	0.23	0.35	0.36
7	0.33	0.21	1.7	n.d.	1	0.39

(n.d. = not detected)

Table 8 shows the gene expression pattern of selected molecules that were upregulated at least 2-fold at least once during the time course following treatment with benzo(a)pyrene (B(a)P). Table 9 shows the gene expression pattern of selected molecules that were downregulated at least 2-fold at least once during the time course following treatment with B(a)P.

TABLE 8 Gene expression patterns (x-fold change) of B(a)P-upregulated nucleic acid molecules

SEQ ID NO:	12 hours	1 day	3 days	7 days	14 days	28 days
2	0.5	0.47	1.2	1.1	2.6	0.47
3	1.4	2.1	1.2	1.5	2.7	1.6
5	1.5	1.4	1.2	0.47	2	0.46
6	2.2	1.4	1.4	1.2	2.2	n.d.
7	1.2	2.2	1.4	0.5	0.42	1.1
8	1.6	1.7	1.3	1.3	2	1.1

(n.d. = not detected)

TABLE 9 Gene expression patterns (x-fold change) of B(a)P-downregulated nucleic acid molecules

SEQ ID NO:	12 hours	1 day	3 days	7 days	14 days	28
1	0.37	0.39	0.35	1.4	0.33	1.5
4	0.5	0.26	0.31	0.36	0.47	n.d.

(n.d. = not detected)

Table 10 shows the library abundance of selected molecules that were up- or down-regulated at least once following treatment with various agents. Library abundance of each SEQ ID NO is presented as relative to that library which included the least abundant levels of nucleic acid molecule (SEQ ID NO) present.

TABLE 10 Library abundance (least abundant = 1) patterns of nucleic acid molecules

SEQ ID NO:	Untreated	CLO	FEN	APAP	BaP	CCl_4	HYDR	ANIT	4-AAF
8	4	7	6	3	9	4	1	1	3
9	13	5	6	4	15	5	6	6	2
10	n.d.	1	8	3	n.d.	n.d.	n.d.	1	n.d.
11	5	2	4	8	20	7	10	n.d.	2

(n.d. = not detected)

XI Identification and Analyses of Homologous Molecule in other Organisms

The rat sequences (SEQ ID NOs:1-16) were used to identify additional sequences in the ZOOSEQ and LIFESEQ databases (Incyte Pharmaceuticals) related to rat nucleic acid molecules regulated during toxicological response (SEQ ID NOs:18-47).

The first pass cDNAs, SEQ ID NOs:5, and 60 through 134, were assembled using PHRAP (Phil Green, supra), using the following default parameters, to produce the contiguous sequence SEQ ID NO:135. Mismatch penalty = -2; gap initiation penalty <0; gap extension penalty <0; minimum length of matching word = 14; minimum SWAT score = 30; bandwidth = 14; use raw SW scores, "No"; index word size = 10; maximum gap size = 30; number of initial bases to be converted to 'N', 0; vector segment length = 60; Mismatch penalty for scoring degenerate end sequence = -2; Min. score for converting degenerate end sequence to 'N', 20; Minimum size of confirming segment = 8; Amount by which confirming segments are trimmed = 1; Penalty for confirming matches = -5; Min. SWAT score for confirming matches = 30; LLR cutoff for displaying discrepancies = 20; Minimum segment size = 8; Spacing between nodes = 4; Split/reassemble initial 'greedy' assembly, "No".

Translation of SEQ ID NO:135 using MACDNASIS PRO software (version 1.0, Hitachi Software Engineering) using default parameters of the program elucidated the putative protein coding region, SEQ ID NO:136. The nucleic acid and amino acid sequences were queried against databases such as the LIFESEQ (Incyte), GenBank, and SwissProt databases using BLAST. Motifs, HMM algorithms, and alignments with BLOCKS, PRINTS, Prosite, and PFAM databases were used to perform functional analyses; the antigenic index (Jameson-Wolf analysis) was determined using LASERGENE software (version 1.62d1, DNASTAR). BLAST2 analysis of SEQ ID NOs:135 and 136 using the human EST LIFESEQ database (Incyte) identified Incyte Clone Numbers 746355H1 (SEQ ID NO:137) and 1294663H1 (SEQ ID NO:138) which were assembled with their respective clustered clones to produce SEQ ID NOs:37 and 38 which encoded SEQ ID NOs:51 and 52, respectively.

Functional analysis of SEQ ID NO:136 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential protein kinase C phosphorylation site at residue S84 (Motifs); a potential signal peptide from residue M1 through residue A33 (SPScan); a potential transmembrane domain from residue P37 through residue L56 (HMM TM), a sodium/neurotransmitter symporter signature from residue G34 through A53, a sodium/alanine symporter signature from G34 through A53, and an asparaginase/glutaminase family signature from residue W64 through residue G75 (BOCKS and PRINTS).

Functional analysis of SEQ ID NO:51 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential protein kinase C phosphorylation site at residue S83 (Motifs); a potential signal peptide sequence from residue M1 through residue A52 (SPScan); a sodium/alanine symporter signature from residue G33 through residue A52, an asparaginase/glutaminase family signature from residue W63 through residue G74, and a channel-forming colicin domain from residue K31 through residue G49 (BLOCKS and PRINTS). Functional analysis of SEQ ID NO:52 using BLOCKS, PRINTS, Prosite, PFAM, Motifs, and HMM algorithms identified a potential signal peptide sequence from residue M1 through A53 (SPScan); a sodium/alanine symporter signature from residue G34 through residue A53, a 6-phosphogluconate dehydrogenase family signature from residue G15 through residue A40, an FAD-dependant glycerol-3-phosphate dehydrogenase family signature from residue Y18 through residue Y30, and a vacuolar ATP synthetase 16 kDa subunit signature from residue L39 through residue G65 (BLOCKS and PRINTS).

CLAIMS

What is claimed is:

1. A method for detecting or diagnosing the effect of a toxic compound or molecule associated with increased or decreased levels of nucleic acid molecules in a mammalian subject comprising:

a) treating a mammalian subject with a toxic compound or molecule;

b) obtaining a sample containing nucleic acids from the mammalian subject treated with the toxic compound or molecule;

c) contacting the sample with a microarray comprising a plurality of nucleic acid molecules of SEQ ID NOs:1-47, or a fragment thereof under conditions for the formation of one or more hybridization complexes; and

d) detecting the hybridization complexes, wherein the presence, absence or change in amount of the hybridization complex, as compared with the hybridization complexes formed from nucleic acid molecules from an untreated mammalian subject, is indicative of a metabolic response to the toxic compound or molecule.

2. The method of claim 1 wherein:

a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;

b) the sample is liver tissue;

c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;

d) the toxic compound is a peroxisome proliferator;

e) the toxic compound is a hypolipidemic drug; and

f) the toxic compound is clofibrate or one of its corresponding metabolites.

3. The method of claim 1 wherein:

a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;

b) the sample is liver tissue;

c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;

d) the toxic compound is acetominophen or one of its corresponding metabolites.

4. The method of claim 1 wherein:

a) the sample is a tissue chosen from liver, kidney, brain, spleen, pancreas, and lung;

b) the sample is liver tissue;

c) the toxic compound or molecule which elicits the metabolic response induces at least a 2-fold change in the amount of at least one of the nucleic acid molecules of the sample;

d) the toxic compound is a polycyclic aromatic hydrocarbon;

e) the toxic compound is a diol epoxide; and

f) the toxic compound is benzo(a)pyrene, or one of its corresponding metabolites.

5. A method for detecting or diagnosing a toxicological response to a test compound or molecule in a mammalian subject, the method comprising:

a) treating a mammalian subject with a test compound or molecule;

b) obtaining a sample containing nucleic acids from the mammalian subject treated with the test compound or molecule;

c) contacting the sample with a microarray comprising a plurality of nucleic acid molecules of SEQ ID NOs:1-47, or a fragment thereof, under conditions for the formation of one or more hybridization complexes;

d) detecting the hybridization complexes, wherein the presence, absence or change in amount of the hybridization complex, as compared with the hybridization complexes formed from nucleic acid molecules from a normal or untreated mammalian subject, is indicative of a toxic response to the test compound or molecule.

6. The method of claim 5 wherein the test compound which elicits the metabolic response is a compound with a previously known metabolic response.

7. The method of claim 5 wherein the test compound which elicits the metabolic response is a compound with a previously unknown metabolic response.

8. An isolated and purified nucleic acid molecule selected from SEQ ID NOs:1-11, 17-33, 36, 39, and 41, or a fragment thereof, wherein said fragments are at least 60 contiguous nucleotides in length.

9. A method of using a molecule selected from SEQ ID NOs:1-59 or a fragment thereof to screen a library of molecules or compounds to identify at least one molecule or compound which specifically binds the selected molecule, the method comprising:

a) combining the selected molecule with a library of molecules or compounds under conditions to allow specific binding; and

b) detecting specific binding, thereby identifying a molecule or compound which specifically binds the selected molecule.

10. The method of claim 9 wherein the library is selected from DNA molecules, RNA molecules, peptide nucleic acids, artificial chromosome constructions, peptides, proteins, and drugs.

11. An isolated and purified antibody identified using the method of claim 9.

12. An isolated and purified nucleic acid molecule variant having at least 70% nucleic acid sequence identity to the nucleic acid molecule of claim 8.

13. An isolated and purified nucleic acid molecule having a sequence which is complementary to the nucleic acid molecule of claim 8.

14. An isolated and purified agonist identified using the method of claim 9.

15. An isolated and purified antagonist identified using the method of claim 9.

16. An expression vector comprising at least a fragment of the nucleic acid molecule of claim 8.

17. A host cell comprising the expression vector of claim 16.

18. A method for producing a polypeptide, the method comprising the steps of:

a) culturing the host cell of claim 17 under conditions suitable for the expression of the polypeptide; and

b) recovering the polypeptide from the host cell culture.

19. An isolated and purified protein molecule encoded by the nucleic acid molecule selected from SEQ ID NOs:1-11, 17-33, 36, 39, and 41, an isolated and purified protein molecule of SEQ ID NOs:50 and 53, or a portion thereof, wherein said portions encode at least 20 contiguous amino acids in length.

20. A pharmaceutical composition comprising the protein molecule of claim 19 in conjunction with a suitable pharmaceutical carrier.

SEQUENCE LISTING

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CUNNINGHAM, Mary Jane
ZWEIGER, Gary
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PANZER, Scott
SEILHAMMER, Jeffrey J.
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<211> 264
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<210> 13
<211> 285
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<212> DNA
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<213> Homo sapiens

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<213> Homo sapiens

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<213> Homo sapiens

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<223> Incyte ID No.: 1461451

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<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 2345712

<400> 45

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<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 964996

<400> 47

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<211> 1512
<212> PRT
<213> Homo sapiens

<220>
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<223> Incyte ID No.: 2302721

<400> 48

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Ser Val Glu Glu Gly Lys Glu Asn Ile Leu His Val Ser Glu Asn
             35                  40                  45
Val Ile Phe Thr Asp Val Asn Ser Ile Leu Arg Tyr Leu Ala Arg
             50                  55                  60
Val Ala Thr Thr Ala Gly Leu Tyr Gly Ser Asn Leu Met Glu His
             65                  70                  75
Thr Glu Ile Asp His Trp Leu Glu Phe Ser Ala Thr Lys Leu Ser
             80                  85                  90
Ser Cys Asp Ser Phe Thr Ser Thr Ile Asn Glu Leu Asn His Cys
             95                 100                 105
Leu Ser Leu Arg Thr Tyr Leu Val Gly Asn Ser Leu Ser Leu Ala
            110                 115                 120
Asp Leu Cys Val Trp Ala Thr Leu Lys Gly Asn Ala Ala Trp Gln
            125                 130                 135
Glu Gln Leu Lys Gln Lys Lys Ala Pro Val His Val Lys Arg Trp
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Phe Gly Phe Leu Glu Ala Gln Gln Ala Phe Gln Ser Val Gly Thr
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Lys Trp Asp Val Ser Thr Thr Lys Ala Arg Val Ala Pro Glu Lys
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Lys Gln Asp Val Gly Lys Phe Val Glu Leu Pro Gly Ala Glu Met
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Gly Lys Val Thr Val Arg Phe Pro Pro Glu Ala Ser Gly Tyr Leu
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His Ile Gly His Ala Lys Ala Ala Leu Leu Asn Gln His Tyr Gln
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Val Asn Phe Lys Gly Lys Leu Ile Met Arg Phe Asp Asp Thr Asn
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Pro Glu Lys Glu Lys Glu Asp Phe Glu Lys Val Ile Leu Glu Asp
            245                 250                 255
Val Ala Met Leu His Ile Lys Pro Asp Gln Phe Thr Tyr Thr Ser
            260                 265                 270
Asp His Phe Glu Thr Ile Met Lys Tyr Ala Glu Lys Leu Ile Gln
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Glu Gly Lys Ala Tyr Val Asp Asp Thr Pro Ala Glu Gln Met Lys
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Ala Glu Arg Glu Gln Arg Ile Glu Ser Lys His Arg Lys Asn Pro
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Ile Gln Pro His Pro Arg Thr Gly Asn Lys Tyr Asn Val Tyr Pro
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Thr Tyr Asp Phe Ala Cys Pro Ile Val Asp Ser Ile Glu Gly Val
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Thr His Ala Leu Arg Thr Thr Glu Tyr His Asp Arg Asp Glu Gln
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Phe Tyr Trp Ile Ile Glu Ala Leu Gly Ile Arg Lys Pro Tyr Ile
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Arg Lys Leu Thr Trp Phe Val Asn Glu Gly Leu Val Asp Gly Trp
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Asp Asp Pro Arg Phe Pro Thr Val Arg Gly Val Leu Arg Arg Gly
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Ser Arg Ser Val Val Asn Met Glu Trp Asp Lys Ile Trp Ala Phe
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Asn Lys Lys Val Ile Asp Pro Val Ala Pro Arg Tyr Val Ala Leu
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Leu Lys Lys Glu Val Ile Pro Val Asn Val Pro Glu Ala Gln Glu
                515                 520                 525
Glu Met Lys Glu Val Ala Lys His Pro Lys Asn Pro Glu Val Gly
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Leu Lys Pro Val Trp Tyr Ser Pro Lys Val Phe Ile Glu Gly Ala
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Pro Thr Pro Ser Leu Asn Asn Asn Cys Thr Thr Ser Glu Asp Ser
                740                 745                 750
Leu Val Leu Tyr Asn Arg Val Ala Val Gln Gly Asp Val Val Arg
                755                 760                 765
Glu Leu Lys Ala Lys Lys Ala Pro Lys Glu Asp Val Asp Ala Ala
                770                 775                 780
Val Lys Gln Leu Leu Ser Leu Lys Ala Glu Tyr Lys Glu Lys Thr
                785                 790                 795
Gly Gln Glu Tyr Lys Pro Gly Asn Pro Pro Ala Glu Ile Gly Gln
                800                 805                 810
Asn Ile Ser Ser Asn Ser Ser Ala Ser Ile Leu Glu Ser Lys Ser
```

```
                815                 820                 825
Leu Tyr Asp Glu Val Ala Ala Gln Gly Glu Val Val Arg Lys Leu
                830                 835                 840
Lys Ala Glu Lys Ser Pro Lys Ala Lys Ile Asn Glu Ala Val Glu
                845                 850                 855
Cys Leu Leu Ser Leu Lys Ala Gln Tyr Lys Glu Lys Thr Gly Lys
                860                 865                 870
Glu Tyr Ile Pro Gly Gln Pro Pro Leu Ser Gln Ser Ser Asp Ser
                875                 880                 885
Ser Pro Thr Arg Asn Ser Glu Pro Ala Gly Leu Glu Thr Pro Glu
                890                 895                 900
Ala Lys Val Leu Phe Asp Lys Val Ala Ser Gln Gly Glu Val Val
                905                 910                 915
Arg Lys Leu Lys Thr Glu Lys Ala Pro Lys Asp Gln Val Asp Ile
                920                 925                 930
Ala Val Gln Glu Leu Leu Gln Leu Lys Ala Gln Tyr Lys Ser Leu
                935                 940                 945
Ile Gly Val Glu Tyr Lys Pro Val Ser Ala Thr Gly Ala Glu Asp
                950                 955                 960
Lys Asp Lys Lys Lys Lys Glu Lys Glu Asn Lys Ser Glu Lys Gln
                965                 970                 975
Asn Lys Pro Gln Lys Gln Asn Asp Gly Gln Arg Lys Asp Pro Ser
                980                 985                 990
Lys Asn Gln Gly Gly Gly Leu Ser Ser Ser Gly Ala Gly Glu Gly
                995                1000                1005
Gln Gly Pro Lys Lys Gln Thr Arg Leu Gly Leu Glu Ala Lys Lys
               1010                1015                1020
Glu Glu Asn Leu Ala Asp Trp Tyr Ser Gln Val Ile Thr Lys Ser
               1025                1030                1035
Glu Met Ile Glu Tyr His Asp Ile Ser Gly Cys Tyr Ile Leu Arg
               1040                1045                1050
Pro Trp Ala Tyr Ala Ile Trp Glu Ala Ile Lys Asp Phe Phe Asp
               1055                1060                1065
Ala Glu Ile Lys Lys Leu Gly Val Glu Asn Cys Tyr Phe Pro Met
               1070                1075                1080
Phe Val Ser Gln Ser Ala Leu Glu Lys Glu Lys Thr His Val Ala
               1085                1090                1095
Asp Phe Ala Pro Glu Val Ala Trp Val Thr Arg Ser Gly Lys Thr
               1100                1105                1110
Glu Leu Ala Glu Pro Ile Ala Ile Arg Pro Thr Ser Glu Thr Val
               1115                1120                1125
Met Tyr Pro Ala Tyr Ala Lys Trp Val Gln Ser His Arg Asp Leu
               1130                1135                1140
Pro Ile Lys Leu Asn Gln Trp Cys Asn Val Val Arg Trp Glu Phe
               1145                1150                1155
Lys His Pro Gln Pro Phe Leu Arg Thr Arg Glu Phe Leu Trp Gln
               1160                1165                1170
Glu Gly His Ser Ala Phe Ala Thr Met Glu Glu Ala Ala Glu Glu
               1175                1180                1185
Val Leu Gln Ile Leu Asp Leu Tyr Ala Gln Val Tyr Glu Glu Leu
               1190                1195                1200
Leu Ala Ile Pro Val Val Lys Gly Arg Lys Thr Glu Lys Glu Lys
               1205                1210                1215
Phe Ala Gly Gly Asp Tyr Thr Thr Thr Ile Glu Ala Phe Ile Ser
               1220                1225                1230
Ala Ser Gly Arg Ala Ile Gln Gly Gly Thr Ser His His Leu Gly
               1235                1240                1245
Gln Asn Phe Ser Lys Met Phe Glu Ile Val Phe Glu Asp Pro Lys
               1250                1255                1260
Ile Pro Gly Glu Lys Gln Phe Ala Tyr Gln Asn Ser Trp Gly Leu
               1265                1270                1275
Thr Thr Arg Thr Ile Gly Val Met Thr Met Val His Gly Asp Asn
               1280                1285                1290
Met Gly Leu Val Leu Pro Pro Arg Val Ala Cys Val Gln Val Val
               1295                1300                1305
Ile Ile Pro Cys Gly Ile Thr Asn Ala Leu Ser Glu Glu Asp Lys
               1310                1315                1320
Glu Ala Leu Ile Ala Lys Cys Asn Asp Tyr Arg Arg Arg Leu Leu
```

```
                1325                1330                1335
Ser Val Asn Ile Arg Val Arg Ala Asp Leu Arg Asp Asn Tyr Ser
                1340                1345                1350
Pro Gly Trp Lys Phe Asn His Trp Glu Leu Lys Gly Val Pro Ile
                1355                1360                1365
Arg Leu Glu Val Gly Pro Arg Asp Met Lys Ser Cys Gln Phe Val
                1370                1375                1380
Ala Val Arg Arg Asp Thr Gly Glu Lys Leu Thr Val Ala Glu Asn
                1385                1390                1395
Glu Ala Glu Thr Lys Leu Gln Ala Ile Leu Glu Asp Ile Gln Val
                1400                1405                1410
Thr Leu Phe Thr Arg Ala Ser Glu Asp Leu Lys Thr His Met Val
                1415                1420                1425
Val Ala Asn Thr Met Glu Asp Phe Gln Lys Ile Leu Asp Ser Gly
                1430                1435                1440
Lys Ile Val Gln Ile Pro Phe Cys Gly Glu Ile Asp Cys Glu Asp
                1445                1450                1455
Trp Ile Lys Lys Thr Thr Ala Arg Asp Gln Asp Leu Glu Pro Gly
                1460                1465                1470
Ala Pro Ser Met Gly Ala Lys Ser Leu Cys Ile Pro Phe Lys Pro
                1475                1480                1485
Leu Cys Glu Leu Gln Pro Gly Ala Lys Cys Val Cys Gly Lys Asn
                1490                1495                1500
Pro Ala Lys Tyr Tyr Thr Leu Phe Gly Arg Ser Tyr
                1505                1510
```

<210> 49
<211> 238
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 2742442

<400> 49

```
Met Ala Ala Arg Thr Gly His Thr Ala Leu Arg Arg Val Val Ser
  1               5                  10                  15
Gly Cys Arg Pro Lys Ser Ala Thr Ala Ala Gly Ala Gln Ala Pro
                 20                  25                  30
Val Arg Asn Gly Arg Tyr Leu Ala Ser Cys Gly Ile Leu Met Ser
                 35                  40                  45
Arg Thr Leu Pro Leu His Thr Ser Ile Leu Pro Lys Glu Ile Cys
                 50                  55                  60
Ala Arg Thr Phe Phe Lys Ile Thr Ala Pro Leu Ile Asn Lys Arg
                 65                  70                  75
Lys Glu Tyr Ser Glu Arg Arg Ile Leu Gly Tyr Ser Met Gln Glu
                 80                  85                  90
Met Tyr Asp Val Val Ser Gly Val Glu Asp Tyr Lys His Phe Val
                 95                 100                 105
Pro Trp Cys Lys Lys Ser Asp Val Ile Ser Lys Arg Ser Gly Tyr
                110                 115                 120
Cys Lys Thr Arg Leu Glu Ile Gly Phe Pro Pro Val Leu Glu Arg
                125                 130                 135
Tyr Thr Ser Val Val Thr Leu Val Lys Pro His Leu Val Lys Ala
                140                 145                 150
Ser Cys Thr Asp Gly Arg Leu Phe Asn His Leu Glu Thr Ile Trp
                155                 160                 165
Cys Phe Ser Pro Gly Leu Pro Gly Tyr Pro Arg Thr Cys Thr Leu
                170                 175                 180
Asp Phe Ser Ile Ser Phe Glu Phe Arg Ser Leu Leu His Ser Gln
                185                 190                 195
Leu Ala Thr Leu Phe Phe Asp Glu Val Val Lys Gln Met Val Ala
                200                 205                 210
```

```
Ala Phe Glu Arg Arg Ala Cys Lys Leu Tyr Gly Pro Glu Thr Asn
                215                 220                 225
Ile Pro Arg Glu Leu Met Leu His Glu Val His His Thr
                230                 235
```

<210> 50
<211> 653
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 3511087

<400> 50

```
Met Pro Phe Ser Ala Ser Leu Leu Gly Thr Leu Pro Ile Gly Ala
  1               5                  10                  15
Arg Tyr Ala Pro Pro Pro Ser Phe Ser Glu Phe Tyr Pro Pro Leu
             20                  25                  30
Thr Ser Ser Leu Glu Asp Phe Cys Ser Ser Leu Asn Ser Phe Ser
             35                  40                  45
Met Ser Glu Ser Lys Arg Asp Leu Ser Thr Ser Thr Ser Arg Glu
             50                  55                  60
Gly Thr Pro Leu Asn Asn Ser Asn Ser Ser Leu Leu Leu Met Asn
             65                  70                  75
Gly Pro Gly Ser Leu Phe Ala Ser Glu Asn Phe Leu Gly Ile Ser
             80                  85                  90
Ser Gln Pro Arg Asn Asp Phe Gly Asn Phe Phe Gly Ser Ala Val
             95                 100                 105
Thr Lys Pro Ser Ser Ser Val Thr Pro Arg His Pro Leu Glu Gly
            110                 115                 120
Thr His Glu Leu Arg Gln Ala Cys Gln Ile Cys Phe Val Lys Ser
            125                 130                 135
Gly Pro Lys Leu Met Asp Phe Thr Tyr His Ala Asn Ile Asp His
            140                 145                 150
Lys Cys Lys Lys Asp Ile Leu Ile Gly Arg Ile Lys Asn Val Glu
            155                 160                 165
Asp Lys Ser Trp Lys Lys Ile Arg Pro Arg Pro Thr Lys Thr Asn
            170                 175                 180
Tyr Glu Gly Pro Tyr Tyr Ile Cys Lys Asp Val Ala Ala Glu Glu
            185                 190                 195
Glu Cys Arg Tyr Ser Gly His Cys Thr Phe Ala Tyr Cys Gln Glu
            200                 205                 210
Glu Ile Asp Val Trp Thr Leu Glu Arg Lys Gly Ala Phe Ser Arg
            215                 220                 225
Glu Ala Phe Phe Gly Gly Asn Gly Lys Ile Asn Leu Thr Val Phe
            230                 235                 240
Lys Leu Leu Gln Glu His Leu Gly Glu Phe Ile Phe Leu Cys Glu
            245                 250                 255
Lys Cys Phe Asp His Lys Pro Arg Met Ile Ser Lys Arg Asn Lys
            260                 265                 270
Asp Asn Ser Thr Ala Cys Ser His Pro Val Thr Lys His Glu Phe
            275                 280                 285
Glu Asp Asn Lys Cys Leu Val His Ile Leu Arg Glu Thr Thr Val
            290                 295                 300
Lys Tyr Ser Lys Ile Arg Ser Phe His Gly Gln Cys Gln Leu Asp
            305                 310                 315
Leu Cys Arg His Glu Val Arg Tyr Gly Cys Leu Arg Glu Asp Glu
            320                 325                 330
Cys Phe Tyr Ala His Ser Leu Val Glu Leu Lys Val Trp Ile Met
            335                 340                 345
Gln Asn Glu Thr Gly Ile Ser His Asp Ala Ile Ala Gln Glu Ser
            350                 355                 360
Lys Arg Tyr Trp Gln Asn Leu Glu Ala Asn Val Pro Gly Ala Gln
            365                 370                 375
Val Leu Gly Asn Gln Ile Met Pro Gly Phe Leu Asn Met Lys Ile
```

```
                380                 385                 390
Lys Phe Val Cys Ala Gln Cys Leu Arg Asn Gly Gln Val Ile Glu
                395                 400                 405
Pro Asp Lys Asn Arg Lys Tyr Cys Ser Ala Lys Ala Arg His Ser
                410                 415                 420
Trp Thr Lys Asp Arg Arg Ala Met Arg Val Met Ser Ile Glu Arg
                425                 430                 435
Lys Lys Trp Met Asn Ile Arg Pro Leu Pro Thr Lys Lys Gln Met
                440                 445                 450
Pro Leu Gln Phe Asp Leu Cys Asn His Ile Ala Ser Gly Lys Lys
                455                 460                 465
Cys Gln Tyr Val Gly Asn Cys Ser Phe Ala His Ser Pro Glu Glu
                470                 475                 480
Arg Glu Val Trp Thr Tyr Met Lys Glu Asn Gly Ile Gln Asp Met
                485                 490                 495
Glu Gln Phe Tyr Glu Leu Trp Leu Lys Ser Gln Lys Asn Glu Lys
                500                 505                 510
Ser Glu Asp Ile Ala Ser Gln Ser Asn Lys Glu Asn Gly Lys Gln
                515                 520                 525
Ile His Met Pro Thr Asp Tyr Ala Glu Val Thr Val Asp Phe His
                530                 535                 540
Cys Trp Met Cys Gly Lys Asn Cys Asn Ser Glu Lys Gln Trp Gln
                545                 550                 555
Gly His Ile Ser Ser Glu Lys His Lys Glu Lys Val Phe His Thr
                560                 565                 570
Glu Asp Asp Gln Tyr Cys Trp Gln His Arg Phe Pro Thr Gly Tyr
                575                 580                 585
Phe Ser Ile Cys Asp Arg Tyr Met Asn Gly Thr Cys Pro Glu Gly
                590                 595                 600
Asn Ser Cys Lys Phe Ala His Gly Asn Ala Glu Leu His Glu Trp
                605                 610                 615
Glu Glu Arg Arg Asp Ala Leu Lys Met Lys Leu Asn Lys Ala Arg
                620                 625                 630
Lys Asp His Leu Ile Gly Pro Asn Asp Asn Asp Phe Gly Lys Tyr
                635                 640                 645
Ser Phe Leu Phe Lys Asp Leu Asn
                650
```

<210> 51
<211> 112
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1968009

<400> 51

```
Met Gln Asp Thr Gly Ser Val Val Pro Leu His Trp Phe Gly Phe
  1               5                  10                  15
Gly Tyr Ala Ala Leu Val Ala Ser Gly Gly Ile Ile Gly Tyr Val
                20                  25                  30
Lys Ala Gly Ser Val Pro Ser Leu Ala Ala Gly Leu Leu Phe Gly
                35                  40                  45
Ser Leu Ala Gly Leu Gly Ala Tyr Gln Leu Ser Gln Asp Pro Arg
                50                  55                  60
Asn Val Trp Val Phe Leu Ala Thr Ser Gly Thr Leu Ala Gly Ile
                65                  70                  75
Met Gly Met Arg Phe Tyr His Ser Gly Lys Phe Met Pro Ala Gly
                80                  85                  90
Leu Ile Ala Gly Ala Ser Leu Leu Met Val Ala Lys Val Gly Val
                95                 100                 105
Ser Met Phe Asn Arg Pro His
               110
```

<210> 52
<211> 114
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1923127

<400> 52

```
Met Glu Lys Pro Leu Phe Pro Leu Val Pro Leu His Trp Phe Gly
  1               5                  10                  15
Phe Gly Tyr Thr Ala Leu Val Val Ser Gly Gly Ile Val Gly Tyr
             20                  25                  30
Val Lys Thr Gly Ser Val Pro Ser Leu Ala Ala Gly Leu Leu Phe
             35                  40                  45
Gly Ser Leu Ala Gly Leu Gly Ala Tyr Gln Leu Tyr Gln Asp Pro
             50                  55                  60
Arg Asn Val Trp Gly Phe Leu Ala Ala Thr Ser Val Thr Phe Val
             65                  70                  75
Gly Val Met Gly Met Arg Ser Tyr Tyr Tyr Gly Lys Phe Met Pro
             80                  85                  90
Val Gly Leu Ile Ala Gly Ala Ser Leu Leu Met Ala Ala Lys Val
             95                 100                 105
Gly Val Arg Met Leu Met Thr Ser Asp
            110
```

<210> 53
<211> 114
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 3123954

<400> 53

```
Met Ala Ala Ile Pro Ser Ser Gly Ser Leu Val Ala Thr His Asp
  1               5                  10                  15
Tyr Tyr Arg Arg Arg Leu Gly Ser Thr Ser Ser Asn Ser Ser Cys
             20                  25                  30
Ser Ser Thr Glu Cys Pro Gly Glu Ala Ile Pro His Pro Pro Gly
             35                  40                  45
Leu Pro Lys Ala Asp Pro Gly His Trp Trp Ala Ser Phe Phe Phe
             50                  55                  60
Gly Lys Ser Thr Leu Pro Phe Met Ala Thr Val Leu Glu Ser Ala
             65                  70                  75
Glu His Ser Glu Pro Pro Gln Ala Ser Ser Ser Met Thr Ala Cys
             80                  85                  90
Gly Leu Ala Arg Asp Ala Pro Arg Lys Gln Pro Gly Gly Gln Ser
             95                 100                 105
Ser Thr Ala Ser Ala Gly Pro Pro Ser
            110
```

<210> 54
<211> 291
<212> PRT
<213> Homo sapiens

<220>
<221>
<223>

<400> 54

```
Met Ser Gln Glu Gly Val Glu Leu Glu Lys Ser Val Arg Gly Leu
  1               5                  10                  15
Arg Glu Lys Phe His Gly Lys Val Ser Ser Lys Lys Ala Gly Ala
             20                  25                  30
Leu Met Arg Lys Phe Gly Ser Asp His Thr Gly Val Gly Arg Ser
             35                  40                  45
Ile Val Tyr Gly Val Lys Gln Lys Asp Gly Gln Glu Leu Ser Asn
             50                  55                  60
Asp Leu Asp Ala Gln Asp Pro Pro Glu Asp Met Lys Gln Asp Arg
             65                  70                  75
Asp Ile Gln Ala Val Ala Thr Ser Leu Leu Pro Leu Thr Glu Ala
             80                  85                  90
Asn Leu Arg Met Phe Gln Arg Ala Gln Asp Asp Leu Ile Pro Ala
             95                 100                 105
Val Asp Arg Gln Phe Ala Cys Ser Ser Cys Asp His Val Trp Trp
            110                 115                 120
Arg Arg Val Pro Gln Arg Lys Glu Val Ser Arg Cys Arg Lys Cys
            125                 130                 135
Arg Lys Arg Tyr Glu Pro Val Pro Ala Asp Lys Met Trp Gly Leu
            140                 145                 150
Ala Glu Phe His Cys Pro Lys Cys Arg His Asn Phe Arg Gly Trp
            155                 160                 165
Ala Gln Met Gly Ser Pro Ser Pro Cys Tyr Gly Cys Gly Phe Pro
            170                 175                 180
Val Tyr Pro Thr Arg Ile Leu Pro Pro Arg Trp Asp Arg Asp Pro
            185                 190                 195
Asp Arg Arg Ser Thr His Thr His Ser Cys Ser Ala Ala Asp Cys
            200                 205                 210
Tyr Asn Arg Arg Glu Pro His Val Pro Gly Thr Ser Cys Ala His
            215                 220                 225
Pro Lys Ser Arg Lys Gln Asn His Leu Pro Lys Val Leu His Pro
            230                 235                 240
Ser Asn Pro His Ile Ser Ser Gly Ser Thr Val Ala Thr Cys Leu
            245                 250                 255
Ser Gln Gly Gly Leu Leu Glu Asp Leu Asp Asn Leu Ile Leu Glu
            260                 265                 270
Asp Leu Lys Glu Glu Glu Glu Glu Glu Glu Glu Val Glu Asp Glu
            275                 280                 285
Glu Gly Gly Pro Arg Glu
            290
```

<210> 55
<211> 610
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1867333

<400> 55

```
Met Trp Leu Pro Leu Val Leu Leu Leu Ala Val Leu Leu Leu Ala
  1               5                  10                  15
Val Leu Cys Lys Val Tyr Leu Gly Leu Phe Ser Gly Ser Ser Pro
             20                  25                  30
Asn Pro Phe Ser Glu Asp Val Lys Arg Pro Pro Ala Pro Leu Val
             35                  40                  45
Thr Asp Lys Glu Ala Arg Lys Lys Val Leu Lys Gln Ala Phe Ser
             50                  55                  60
Ala Asn Gln Val Pro Glu Lys Leu Asp Val Val Val Ile Gly Ser
             65                  70                  75
Gly Phe Gly Gly Leu Ala Ala Ala Ala Ile Leu Ala Lys Ala Gly
             80                  85                  90
Lys Arg Val Leu Val Leu Glu Gln His Thr Lys Ala Gly Gly Cys
```

```
                 95                  100                 105
Cys His Thr Phe Gly Lys Asn Gly Leu Glu Phe Asp Thr Gly Ile
                110                 115                 120
His Tyr Ile Gly Arg Met Glu Glu Gly Ser Ile Gly Arg Phe Ile
                125                 130                 135
Leu Asp Gln Ile Thr Glu Gly Gln Leu Asp Trp Ala Pro Leu Ser
                140                 145                 150
Ser Pro Phe Asp Ile Met Val Leu Glu Gly Pro Asn Gly Arg Lys
                155                 160                 165
Glu Tyr Pro Met Tyr Ser Gly Glu Lys Ala Tyr Ile Gln Gly Leu
                170                 175                 180
Lys Glu Lys Phe Pro Gln Glu Glu Ala Ile Ile Asp Lys Tyr Ile
                185                 190                 195
Lys Leu Val Lys Val Val Ser Ser Gly Ala Pro His Ala Ile Leu
                200                 205                 210
Leu Lys Phe Leu Pro Leu Pro Val Val Gln Leu Leu Asp Arg Cys
                215                 220                 225
Gly Leu Leu Thr Arg Phe Ser Pro Phe Leu Gln Ala Ser Thr Gln
                230                 235                 240
Ser Leu Ala Glu Val Leu Gln Gln Leu Gly Ala Ser Ser Glu Leu
                245                 250                 255
Gln Ala Val Leu Ser Tyr Ile Phe Pro Thr Tyr Gly Val Thr Pro
                260                 265                 270
Asn His Ser Ala Phe Ser Met His Ala Leu Leu Val Asn His Tyr
                275                 280                 285
Met Lys Gly Gly Phe Tyr Pro Arg Gly Gly Ser Ser Glu Ile Ala
                290                 295                 300
Phe His Thr Ile Pro Val Ile Gln Arg Ala Gly Gly Ala Val Leu
                305                 310                 315
Thr Lys Ala Thr Val Gln Ser Val Leu Leu Asp Ser Ala Gly Lys
                320                 325                 330
Ala Cys Gly Val Ser Val Lys Lys Gly His Glu Leu Val Asn Ile
                335                 340                 345
Tyr Cys Pro Ile Val Val Ser Asn Ala Gly Leu Phe Asn Thr Tyr
                350                 355                 360
Glu His Leu Leu Pro Gly Asn Ala Arg Cys Leu Pro Gly Val Lys
                365                 370                 375
Gln Gln Leu Gly Thr Val Arg Pro Gly Leu Gly Met Thr Ser Val
                380                 385                 390
Phe Ile Cys Leu Arg Gly Thr Lys Glu Asp Leu His Leu Pro Ser
                395                 400                 405
Thr Asn Tyr Tyr Val Tyr Tyr Asp Thr Asp Met Asp Gln Ala Met
                410                 415                 420
Glu Arg Tyr Val Ser Met Pro Arg Glu Glu Ala Ala Glu His Ile
                425                 430                 435
Pro Leu Leu Phe Phe Ala Phe Pro Ser Ala Lys Asp Pro Thr Trp
                440                 445                 450
Glu Asp Arg Phe Pro Gly Arg Ser Thr Met Ile Met Leu Ile Pro
                455                 460                 465
Thr Ala Tyr Glu Trp Phe Glu Glu Trp Gln Ala Glu Leu Lys Gly
                470                 475                 480
Lys Arg Gly Ser Asp Tyr Glu Thr Phe Lys Asn Ser Phe Val Glu
                485                 490                 495
Ala Ser Met Ser Val Val Leu Lys Leu Phe Pro Gln Leu Glu Gly
                500                 505                 510
Lys Val Glu Ser Val Thr Ala Gly Ser Pro Leu Thr Asn Gln Phe
                515                 520                 525
Tyr Leu Ala Ala Pro Arg Gly Ala Cys Tyr Gly Ala Asp His Asp
                530                 535                 540
Leu Gly Arg Leu His Pro Cys Val Met Ala Ser Leu Arg Ala Gln
                545                 550                 555
Ser Pro Ile Pro Asn Leu Tyr Leu Thr Gly Gln Asp Ile Phe Thr
                560                 565                 570
Cys Gly Leu Val Gly Ala Leu Gln Gly Ala Leu Leu Cys Ser Ser
                575                 580                 585
Ala Ile Leu Lys Arg Asn Leu Tyr Ser Asp Leu Lys Asn Leu Asp
                590                 595                 600
Ser Arg Ile Arg Ala Gln Lys Lys Lys Asn
```

605 610

<210> 56
<211> 352
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1461451

<400> 56

Pro Arg Val Arg Gly Arg Trp Val Ala His Ala Ser Ala His Ala
1 5 10 15
Ser Ala His Ala Ser Asp Glu Ile Pro Ala Ser Gly Trp Thr Gln
20 25 30
Trp Phe Cys Thr Glu Ala Leu Val Met Val Ala Pro Val Trp Tyr
35 40 45
Leu Val Ala Ala Ala Leu Leu Val Gly Phe Ile Leu Phe Leu Thr
50 55 60
Arg Ser Arg Gly Arg Ala Ala Ser Ala Gly Gln Glu Pro Leu His
65 70 75
Asn Glu Glu Leu Ala Gly Ala Gly Arg Val Ala Gln Pro Gly Pro
80 85 90
Leu Glu Pro Glu Glu Pro Arg Ala Gly Gly Arg Pro Arg Arg Arg
95 100 105
Arg Asp Leu Gly Ser Arg Leu Gln Ala Gln Arg Arg Ala Gln Arg
110 115 120
Val Ala Trp Ala Glu Ala Asp Glu Asn Glu Glu Glu Ala Val Ile
125 130 135
Leu Ala Gln Glu Glu Glu Gly Val Glu Lys Pro Ala Glu Thr His
140 145 150
Leu Ser Gly Lys Ile Gly Ala Lys Lys Leu Arg Lys Leu Glu Glu
155 160 165
Lys Gln Ala Arg Lys Ala Gln Arg Glu Ala Glu Glu Ala Glu Arg
170 175 180
Glu Glu Arg Lys Arg Leu Glu Ser Gln Arg Glu Ala Glu Trp Lys
185 190 195
Lys Glu Glu Glu Arg Leu Arg Leu Glu Glu Glu Gln Lys Glu Glu
200 205 210
Glu Glu Arg Lys Ala Arg Glu Glu Gln Ala Gln Arg Glu His Glu
215 220 225
Glu Tyr Leu Lys Leu Lys Glu Ala Phe Val Val Glu Glu Glu Gly
230 235 240
Val Gly Glu Thr Met Thr Glu Glu Gln Ser Gln Ser Phe Leu Thr
245 250 255
Glu Phe Ile Asn Tyr Ile Lys Gln Ser Lys Val Val Leu Leu Glu
260 265 270
Asp Leu Ala Ser Gln Val Gly Leu Arg Thr Gln Asp Thr Ile Asn
275 280 285
Arg Ile Gln Asp Leu Leu Ala Glu Gly Thr Ile Thr Gly Val Ile
290 295 300
Asp Asp Arg Gly Lys Phe Ile Tyr Ile Thr Pro Glu Glu Leu Ala
305 310 315
Ala Val Ala Asn Phe Ile Arg Gln Arg Gly Arg Val Ser Ile Ala
320 325 330
Glu Leu Ala Gln Ala Ser Asn Ser Leu Ile Ala Trp Gly Arg Glu
335 340 345
Ser Pro Ala Gln Ala Pro Ala
350

<210> 57
<211> 216
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 2345712

<400> 57

```
Tyr Asp Pro Ile Gly Phe Gly Leu Ser Trp Glu Ala Gly Arg Ile
  1               5                  10                  15
Ile Gly Trp Gly Lys Pro Thr Arg Gly Arg Gly Arg Gly Gly Ser
             20                  25                  30
Leu Ser Thr Arg Gly Arg Gly Ser Glu Val Pro Asp Ser Ala His
             35                  40                  45
Leu Ala Pro Thr Pro Leu Phe Ser Glu Ser Gly Cys Cys Gly Leu
             50                  55                  60
Arg Ser Arg Phe Leu Thr Asp Cys Lys Met Glu Glu Gly Gly Asn
             65                  70                  75
Leu Gly Gly Leu Ile Lys Met Val His Leu Leu Val Leu Ser Gly
             80                  85                  90
Ala Trp Gly Met Gln Met Trp Val Thr Phe Val Ser Gly Phe Leu
             95                 100                 105
Leu Phe Arg Ser Leu Pro Arg His Thr Phe Gly Leu Val Gln Ser
            110                 115                 120
Lys Leu Phe Pro Phe Tyr Phe His Ile Ser Met Gly Cys Ala Phe
            125                 130                 135
Ile Asn Leu Cys Ile Leu Ala Ser Gln His Ala Trp Ala Gln Leu
            140                 145                 150
Thr Phe Trp Glu Ala Ser Gln Leu Tyr Leu Leu Phe Leu Ser Leu
            155                 160                 165
Thr Leu Ala Thr Val Asn Ala Arg Trp Leu Glu Pro Arg Thr Thr
            170                 175                 180
Ala Ala Met Trp Ala Leu Gln Thr Val Glu Lys Glu Arg Gly Leu
            185                 190                 195
Gly Gly Glu Val Pro Gly Ser His Gln Gly Ser Asp Pro Tyr Arg
            200                 205                 210
Gln Leu Arg Glu Lys Asp
            215
```

<210> 58
<211> 292
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1810320

<400> 58

```
Met Ala Gln Pro Pro Pro Asp Val Glu Gly Asp Asp Cys Leu Pro
  1               5                  10                  15
Ala Tyr Arg His Leu Phe Cys Pro Asp Leu Leu Arg Asp Lys Val
             20                  25                  30
Ala Phe Ile Thr Gly Gly Gly Ser Gly Ile Gly Phe Arg Ile Ala
             35                  40                  45
Glu Ile Phe Met Arg His Gly Cys His Thr Val Ile Ala Ser Arg
             50                  55                  60
Ser Leu Pro Arg Val Leu Thr Ala Ala Arg Lys Leu Ala Gly Ala
             65                  70                  75
Thr Gly Arg Arg Cys Leu Pro Leu Ser Met Asp Val Arg Ala Pro
             80                  85                  90
Pro Ala Val Met Ala Ala Val Asp Gln Ala Leu Lys Glu Phe Gly
             95                 100                 105
Arg Ile Asp Ile Leu Ile Asn Cys Ala Ala Gly Asn Phe Leu Cys
            110                 115                 120
Pro Ala Gly Ala Leu Ser Phe Asn Ala Phe Lys Thr Val Met Asp
            125                 130                 135
Ile Asp Thr Ser Gly Thr Phe Asn Val Ser Arg Val Leu Tyr Glu
```

```
                140                 145                 150
Lys Phe Phe Arg Asp His Gly Gly Val Ile Val Asn Ile Thr Ala
                155                 160                 165
Thr Leu Gly Asn Arg Gly Gln Ala Leu Gln Val His Ala Gly Ser
                170                 175                 180
Ala Lys Ala Ala Val Asp Ala Met Thr Arg His Leu Ala Val Glu
                185                 190                 195
Trp Gly Pro Gln Asn Ile Arg Val Asn Ser Leu Ala Pro Gly Pro
                200                 205                 210
Ile Ser Gly Thr Glu Gly Leu Arg Arg Leu Gly Gly Pro Gln Ala
                215                 220                 225
Ser Leu Ser Thr Lys Val Thr Ala Ser Pro Leu Gln Arg Leu Gly
                230                 235                 240
Asn Lys Thr Glu Ile Ala His Ser Val Leu Tyr Leu Ala Ser Pro
                245                 250                 255
Leu Ala Ser Tyr Val Thr Gly Ala Val Leu Val Ala Asp Gly Gly
                260                 265                 270
Ala Trp Leu Thr Phe Pro Asn Gly Val Lys Gly Leu Pro Asp Phe
                275                 280                 285
Ala Ser Phe Ser Ala Lys Leu
                290
```

<210> 59
<211> 158
<212> PRT
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 964996

<400> 59

```
Glu Gly Gly Pro Ser Trp Thr Arg Glu Arg Thr Leu Val Ala Val
  1               5                  10                  15
Lys Pro Asp Gly Val Gln Arg Arg Leu Val Gly Asp Val Ile Gln
                 20                  25                  30
Arg Phe Glu Arg Arg Gly Phe Thr Leu Val Gly Met Lys Met Leu
                 35                  40                  45
Gln Ala Pro Glu Ser Val Leu Ala Glu His Tyr Gln Asp Leu Arg
                 50                  55                  60
Arg Lys Pro Phe Tyr Pro Ala Leu Ile Arg Tyr Met Ser Ser Gly
                 65                  70                  75
Pro Val Val Ala Met Val Trp Glu Gly Tyr Asn Val Val Arg Ala
                 80                  85                  90
Ser Arg Ala Met Ile Gly His Thr Asp Ser Ala Glu Ala Ala Pro
                 95                 100                 105
Gly Thr Ile Arg Gly Tyr Phe Ser Val His Ile Ser Arg Asn Val
                110                 115                 120
Ile His Ala Ser Asp Ser Val Glu Gly Ala Gln Arg Glu Ile Gln
                125                 130                 135
Leu Trp Phe Gln Ser Ser Glu Leu Val Ser Trp Ala Asp Gly Gly
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Gln His Ser Ser Ile His Pro Ala
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<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701884305H1

<400> 60

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tggtttcggc tatgcggccc tggtggctac tggtgggatt attggctatg caaaagcagg 180
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ggttactata gtgaccgtc 559

<210> 61
<211> 326
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 61

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acgtcctgta actgcggttc cgggagtgga aacctaaacg cgcgtgcgct tcttccacgc 180
cacggaaacc gtgcaggcct ggtgtggtct ccaaagtgac tgaacaatgc agaaggacag 240
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<212> DNA
<213> Rattus norvegicus

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<210> 63
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<212> DNA
<213> Rattus norvegicus

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<400> 63

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tgcaaaagca ggtagtgt 318

<210> 64
<211> 315
<212> DNA
<213> Rattus norvegicus

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cgtccctggc tgctggactc ttctttgggg gcctggcagg cctgggtgcc taccagctgt 240
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ggggatgaga ttcta 315

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<212> DNA
<213> Rattus norvegicus

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taccagctgt ctcaggaccc caggaacgtg tgggttttcc tagctacgtc tgggactttg 240
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gcgggancat ttt 313

<210> 66
<211> 304
<212> DNA
<213> Rattus norvegicus

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<400> 66

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tattatggtt tcggctatgc ggccctggtg gctactggtg ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgctgga ctcttctttg ggggcctggc aggcctgggt 240
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ttgg 304

<210> 67
<211> 327
<212> DNA
<213> Rattus norvegicus

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gtgggattat tgnctttgca aaagcaggta gtgtgccgtc cctggctgtt ggactcttct 180
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ggaaatttat gcctgcagtt tgatcgc 327

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<211> 305
<212> DNA
<213> Rattus norvegicus

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caggtagtgt gccgtccctg gctgctggac tcttctttgg gggcctggca ggcctgggtg 240
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gctgg 305

<210> 69
<211> 295
<212> DNA
<213> Rattus norvegicus

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<223> Incyte ID No.: 700275094H1

<400> 69

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tttcggctat gcggccctgg tggctactgg tgggattatt ggctatgcaa aagcaggtag 180
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<210> 71
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<212> DNA
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<220>
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tttcggctat gcggccctgg tggctactgg tgggattatt ggctatgcaa aagcaggtag 180
tgtgccgtcc ctggctgctg gactcttctt tgggggcctg gcaggcctgg gtgcctacca 240
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<210> 72
<211> 282
<212> DNA
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tattatggtt tcggctatgc ggccctggtg gctactggtg ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgctgga ctcttctttg ggggcctggc aggcctgggt 240
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<210> 73
<211> 281
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700300118H1

<400> 73

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tattatggtt tcggctatgc ggccctggtg gctactggtg ggattattgg ctatgcaaaa 180
gcaggtagtg tgccgtccct ggctgctgga ctcttctttg ggggcctggc aggcctgggt 240
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<210> 74
<211> 292
<212> DNA
<213> Rattus norvegicus

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agtggcccac tggttccttt acattattat ggtttcggct atgcggccct ggtggctact 180
ggtgggatta ttggctatgc aaaagcaggt agtgtgccgt ccctggctgc tggactcttc 240
tttgggggcc tggcaggcct gggtgcctac cagctgtctc aggaccccag ga 292

<210> 75
<211> 289
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700064344H1

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gagctagcgg actgcccgcc tctctaaaac gtcctgtaac tgcggttccg ggagtggaaa 180
cctaaacgcg cgtgcgcttc ttccacgcca cggaaaccgt gcaggcctgg tgtggtctcc 240
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<211> 276
<212> DNA
<213> Rattus norvegicus

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<223> Incyte ID No.: 701423273H1

<400> 76

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<211> 293
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<213> Rattus norvegicus

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<400> 77

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<210> 78
<211> 274
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701462776H1

<400> 78

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ttacattatt atggtttcgg ctatgcggcc ctggtggcta ctggtgggat tattggctat 180
gcaaaagcag gtagtgtgcc gtccctggct gctggactct tctttggggg cctggcaggc 240
ctgggtgcct accagctgtc tcaggacccc agga 274

<210> 79
<211> 282
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700916803H1

<400> 79

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tctcccgtgc actctctgct gagctagcgg actgcccgcc tctctaaaac gtcctgtaac 180
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<210> 80
<211> 280
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700478141H1

<400> 80

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ttatggtttc ggctatgcgg ccctggtggc tactggtggg attattggct atgcaaaagc 180
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<210> 81
<211> 299
<212> DNA
<213> Rattus norvegicus

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<223> Incyte ID No.: 701646690H1

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<210> 82
<211> 286
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701624261H1

<400> 82

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ggtgggatta ttggctatgc aaaagcaggt agtgtgccgt ccctggctgc nngactcttc 180
tttgggggcc tggcaggcct gggtgcctac cagctgtctc aggaccccag gaacgtgtgg 240
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<210> 83
<211> 266
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 83

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tggactcttc tttgggggcc tggcaggcct gggtgcctac cagctgtctc aggaccccag 180
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<211> 262
<212> DNA
<213> Rattus norvegicus

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atgagattct acaactctgg gaaatttatg cctgcaggtt tgatcgcggg agccagtttg 180
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ccctgcgtgg gctcatgatg ag 262

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<212> DNA
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<400> 85

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<212> DNA
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tgtctcagga ccccaggaac gtgtgggt 268

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<212> DNA
<213> Rattus norvegicus

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<210> 88
<211> 262
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 88

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<210> 89
<211> 349
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 89

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ttcttccacg ccacggaaac cgtgcaggcc tggtgtggtc tccaaagtga ctgaacaatg 180
cagaaggaca gtggcccact ggttccttta cattattatg gtttcggcta tgcggccctg 240
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<210> 90
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<212> DNA
<213> Rattus norvegicus

<220>
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<400> 90

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<212> DNA
<213> Rattus norvegicus

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<400> 91

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<210> 92
<211> 273
<212> DNA
<213> Rattus norvegicus

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<223> Incyte ID No.: 701736525H2

<400> 92

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ccctggtggc tactggtggg attattggct atgcaaaagc aggtagtgtg ccgtccctgg 180
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<210> 93
<211> 262
<212> DNA
<213> Rattus norvegicus

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<400> 93

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ttattatggt ttcggctatg cggccctggt ggctactggt gggattattg gctatgcaaa 180
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tgcctaccag ctgtctcagg ac 262

<210> 94
<211> 247
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700181220H1

<400> 94

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cagtggccca ctggttcctt tacattatta tggtttcggc tatgcggccc tggtggctac 180
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<210> 95
<211> 284
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701462707H1

<400> 95

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atgcagaagg acagtggccc actggttcct ttacattatt atggtttcgg ctatgcggcc 180
ctggtggcta ctggtgggat tattggctat gcaaaagcag gtagtgtgcc gtccctggct 240
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<210> 96
<211> 282
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701462863H1

<400> 96

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<210> 97
<211> 281
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701481465H1

<400> 97

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gcggccctgg tggctactgg tgggattatt ggctatgcaa aagcaggtag tgtgccgtcc 180
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<210> 98
<211> 265
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701308467H1

<400> 98

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tggttccttt acattattat ggtttcggct atgcggccct ggtggctact ggtgggatta 180
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tgnagnctgg gtgcctacca gctgt 265

<210> 99
<211> 291
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701564368H1

<400> 99

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aaatttatgc ctgcaggttt gatcgcggga gccagtttgc tgatggttgc caaacttgga 180
cttagtatgt tgagttcacc ccatccgtag tagccatagt cctgcgtggg ctcatgatga 240
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<210> 100
<211> 271
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700533180H1

<400> 100

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ctttggctgg cattatgggg atgagattct acaactctgg gaaatttatg cctgcaggtt 180
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<210> 101
<211> 255
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700124647H1

<400> 101

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ttatggtttc ggctatgcgg ccctggtggc tactggtggg attattggct atgcaaaagc 180
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tannagctgt ctcaa 255

<210> 102
<211> 297
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700537020H1

<400> 102

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ccccaggaac gtgtgggttt tcctagctac tctggaccnt nggctggcat tatggggatg 180
agattctaca actctgggaa atttatgcct gcaggtttga tcgcgggagc cagtttgctg 240
atggttgcca aacttggact tagtatgttg agttcacccc atccgtagta gccatag 297

<210> 103
<211> 261
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700765205H1

<400> 103

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gcccactggt tcctttacat tattatggtt tcggctatgc ggccctggtg gctactggtg 180
ggattattgg ctatgcaaaa gcaggtagtg tgccgtccct ggctgctgga ctcttctttg 240
ggggcctggc aggctgggtg c 261

<210> 104
<211> 312
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701942992H1

<400> 104

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ctgaacaatg cagaaggaca gtggcccact ggttccttta cattattatg gtttcggcta 180
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ncccaggnan nt 312

<210> 105
<211> 241
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701197694H1

<400> 105

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ttacattatt atggtttcgg ctatgcggcc ctggtggcta ctggtgggat tattggctat 180
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g 241

<210> 106
<211> 268
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701024952H1

<400> 106

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aaggncagtg gcccactggt tcctttacat tattatggtt tcggctatgc ggccctggtg 180
gctactggtg ggattattgg ctatgcaaaa gcaggtagtg tgccgtccct ggctgctgga 240
ctctnctttn ggggcctggc aggcttag 268

<210> 107
<211> 318
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701582676H1

<400> 107

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atcgcgggag ccagtttgct gatggttgcc aaacntggac ttagtatgtt gagttcaccc 180
catccgtagt agccatagtc ctgcgtgggc tcatgatgag ttgacactct ccagtcctcc 240
acattaccac gctgaagaga taagaacagc aaagacctac actgagcaca tggaggcgaa 300
gacgtggtta ctatagtg 318

<210> 108
<211> 255
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701293154H1

<400> 108

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tttcctagct acgtctggga ctttggcttg cattatgggg atgagattct acaactctgg 180
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attagtatgt tgagg 255

<210> 109
<211> 254
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701298824H1

<400> 109

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tcggctatgc ggccctggtg gctactggtg ggattattgg ctatgcaaaa gcnggtagtg 180
tgccgccctg gctgctggac tcttctttgg gggcctgcag nctgggtgcc taccagctgt 240
ctcaggaccc agga 254

<210> 110
<211> 294
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700524204H1

<400> 110

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tttgctgatg gttgccaaac ttggacttag tatgttgagt tcaccccatc cgtagtagcc 180
atagccctgc gtgggctcat gatgagttga cactctccag tcctctacat taccacgctg 240
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<210> 111
<211> 289
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700067537H1

<400> 111

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tgaacaatgc agaaggacag tggcccactg gttcctttac attattatgg tttcggctat 180
gcggccctgg tggctactgg tgggattatt ggctatgcaa aagcagtagt gtgccgtccc 240
tggctgctgg atcttctttg ggggctggca ggctgggtgc ctacaactg 289

<210> 112
<211> 276
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 701258019H1

<400> 112

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actggttcct ttacattatt atggnttccg gctatgcggc cctggtggct actggtgnga 180
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<210> 113
<211> 254
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700532493H1

<400> 113

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tttgctgatg gttgccaaac ttggacttag tatgttgagt tcaccccatc cgtagtagcc 180
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gagatanaac agca 254

<210> 114
<211> 282
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700523302H1

<400> 114

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gggtanangn ncccgnnnng nnaagggggn atnttgnnnt acgnaagagc ngntagtgtg 180
ccgtccctgg ctgctggact cttctttggg ggcctggcag gcctgggtgc ctaccagctg 240
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<210> 115
<211> 256
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 701242719H1

<400> 115

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gcagaaggac agtggcccac tggttccttt acattattat ggtttcggct atgcggccct 180
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actcttcttt ggggcc 256

<210> 116
<211> 244
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 116

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tgcctaccag ctgctcagga ccccaggaac gtgtgggttt tcctagctac gtctgggact 180
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cgcg 244

<210> 117
<211> 262
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
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<400> 117

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gaacaatgca gaaggacagt ggcccactgg ttcctttaca ttattatggt ttcggctatc 180
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<210> 118
<211> 261
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 118

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ggnnttggnc canaaangga a 261

<210> 119
<211> 265
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700533285H1

<400> 119

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tggcccactg gttcctttac attattatgg tttcggctat gcggccctgg tggctactgg 180
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gggggcctgg caggcctggg tgcct 265

<210> 120
<211> 247
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700920823H1

<400> 120

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gctatgcggc cctggtggct actggtggga ttattgctat gcaaaagcag gtagtctgcc 240
gctccct 247

<210> 121
<211> 263
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700627607H1

<400> 121

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<210> 122
<211> 265
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700437944H2

<400> 122

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tgntgggatt attggctatn caaaagcagg tagtgtnccg tccctggctg ctggactctt 180
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<210> 123
<211> 343
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 123

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gtagtagcat ancctgcgtg ggctcagatg agtnacactc tccaggcctc cacatttacc 240
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gacgttcaag acgcgatgnt gactcagact ncntgctcat cgg 343

<210> 124
<211> 241
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 124

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g 241

<210> 125
<211> 155
<212> DNA
<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700916103H1

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<210> 126
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<212> DNA
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<400> 126

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agcgg 185

<210> 127
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<212> DNA
<213> Rattus norvegicus

<220>
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<400> 127

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aactc 125

<210> 128
<211> 266
<212> DNA
<213> Rattus norvegicus

<220>
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gacactctcc agtcctctac attaccacgc tgaagagata agaacagcaa agacctacac 180
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<213> Rattus norvegicus

<220>
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<223> Incyte ID No.: 700325006H1

<400> 129

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<210> 130
<211> 263
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 130

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gacctctcct ccacaggtgc aggcctggtg tggnctccaa agtgactgaa caatgcagaa 180
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<210> 131
<211> 258
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700627187H1

<400> 131

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ttgacactct ccagtcctcc acattaccac gctgaagaga taagaacagc aaagacctac 180
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<210> 132
<211> 272
<212> DNA
<213> Rattus norvegicus

<220>
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<400> 132

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0ttaccacgct gaagagatan gaacagcaaa gacctacact gagcacatgg aggcgaagac 180
gtggttacta tagtgaccgt tcagagacgg cgagtgtctg acctcagagc tcacactgct 240
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<210> 133
<211> 253
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700594190H1

<400> 133

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acgtggttac tatagtgacc gttcagagac ggcgagtgtc tgacctcaga gctcacactg 180
ccttcatgcg gcttgttctt gtgtcatgat gtctcgactc tctgtactac tacataaagg 240
ggtaaaatgt tgg 253

<210> 134
<211> 267
<212> DNA
<213> Rattus norvegicus

<220>
<221> misc_feature
<223> Incyte ID No.: 700627108H1

<400> 134

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tgnnttgaac actctccagt cagtccagat naacgncgct gntagagatn aagaccagcn 180
aagacctaca ctgagcacca tggaggcgaa gacgtggtta ctataagtga ccgttcagag 240
acggcgngtg tntggatcan agatcca 267

<210> 135
<211> 650
<212> DNA
<213> Rattus norvegicus

<220>
<223> RnAUG.conN

<400> 135

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aggcctgggt gcctaccagc tgtctcagga ccccaggaac gtgtgggttt tcctagctac 480
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tgcaggtttg atcgcgggag ccagtttgct gatggttgcc aaacttggac ttagtatgtt 600
gagttcaccc catccgtagt agccatagcc ctgcgtgggc tcatgatgag 650

<210> 136
<211> 114
<212> PRT
<213> Rattus norvegicus

<220>
<223> RnAUG.conP

<400> 136

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20 25 30
Ala Lys Ala Gly Ser Val Pro Ser Leu Ala Ala Gly Leu Phe Phe
35 40 45
Gly Gly Leu Ala Gly Leu Gly Ala Tyr Gln Leu Ser Gln Asp Pro
50 55 60
Arg Asn Val Trp Val Phe Leu Ala Thr Ser Gly Thr Leu Ala Gly
65 70 75
Ile Met Gly Met Arg Phe Tyr Asn Ser Gly Lys Phe Met Pro Ala
80 85 90
Gly Leu Ile Ala Gly Ala Ser Leu Leu Met Val Ala Lys Leu Gly
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Leu Ser Met Leu Ser Ser Pro His Pro
110

<210> 137
<211> 223
<212> DNA
<213> Homo sapiens

<220>
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<400> 137

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gggaatgagg ttctaccact ctggaaaatt catgcctgca ggt 223

<210> 138
<211> 243
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<223> Incyte ID No.: 1294663H1

<400> 138

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aggattaaaa atctgcatct tccactattt tcaatgtatt aagagaaata agtgcagcat 180
ttttgcatct gacattttac ctaaaaaaaa aaagacacca aatttggcgg aggggtggaa 240
aat 0 243